

A multispecies GWAS approach uncovers root responses to phosphate deficiency

Marco Giovannetti
Busch/Dagdass group
2nd Monday Seminar

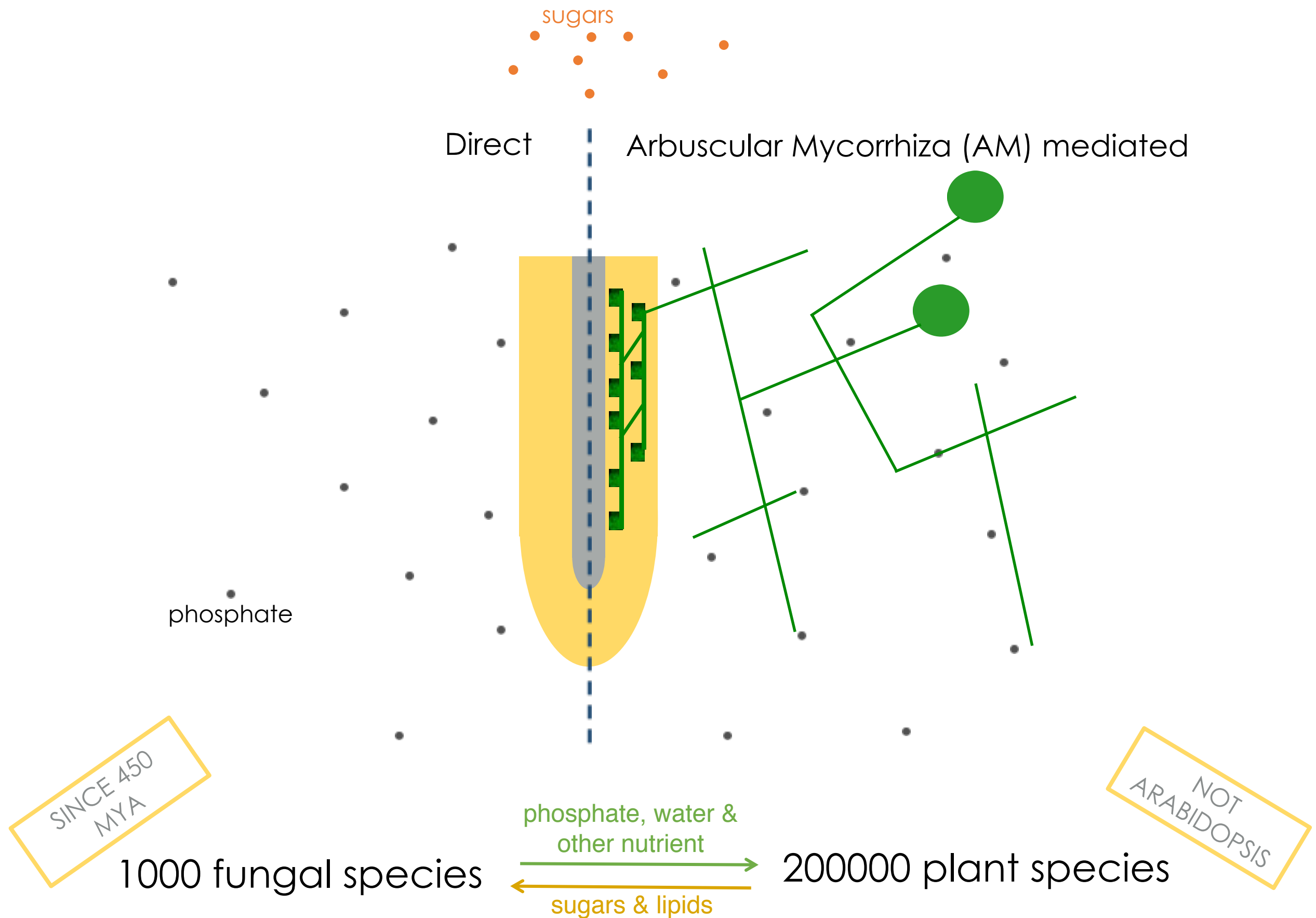
An aerial photograph showing a vast, arid landscape with extensive terraced phosphate mines. The terraces are arranged in a grid-like pattern, creating a series of parallel ridges and valleys. The color of the soil is a deep reddish-brown. In the lower right corner, a small yellow vehicle is visible on one of the terraces.

THE DISAPPEARING NUTRIENT

Nature, 2009

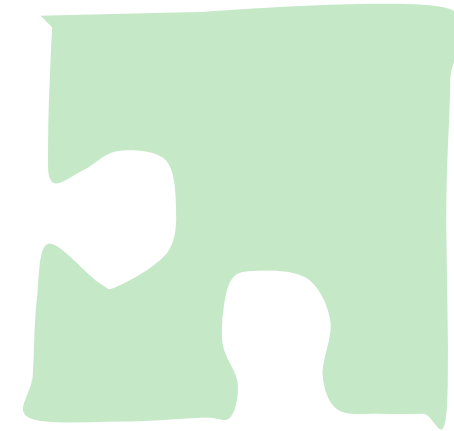
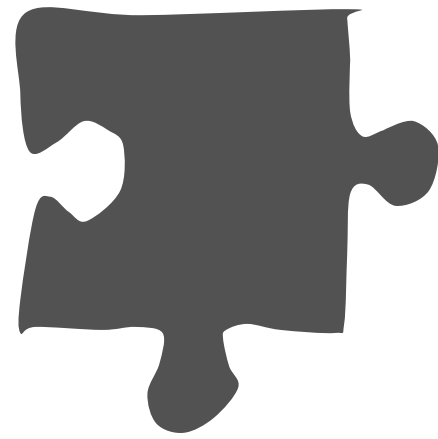
- **Phosphate** is one of the main limiting factors for plant growth
- A limited resource
- Its price is expected to increase year by year
- Much of what is applied is leaching to soil

plant phosphate uptake



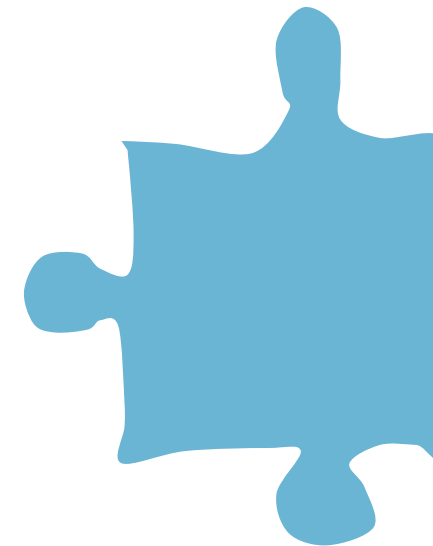
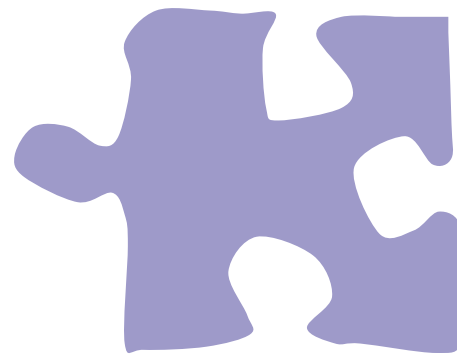
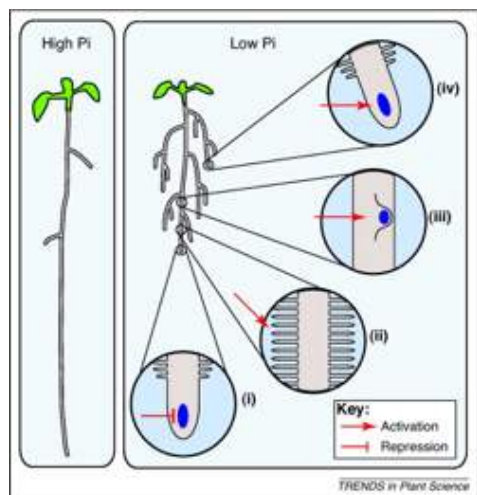
Phosphate puzzle pieces

Rarely
mycorrhizal
phosphate is
considered



Single plant
species
approach

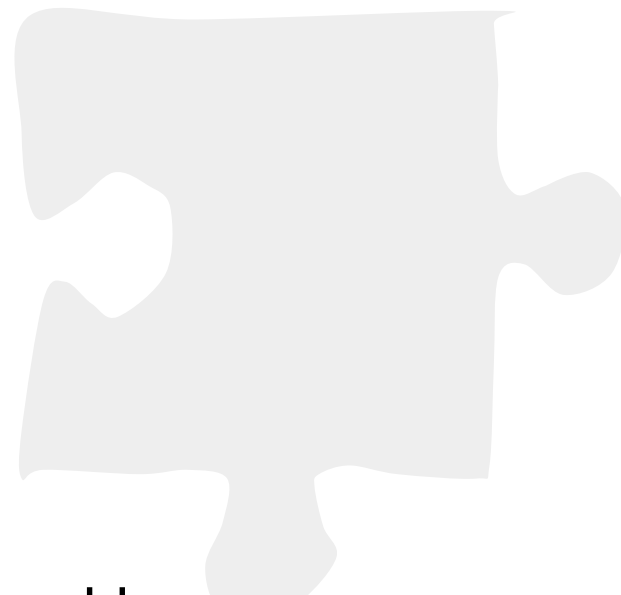
Developmental
responses



Nutritional
changes

Phosphate
transport, recovery
and recycling

Regulation of
mycorrhizal
phosphate
transfer



Single plant
species
approach

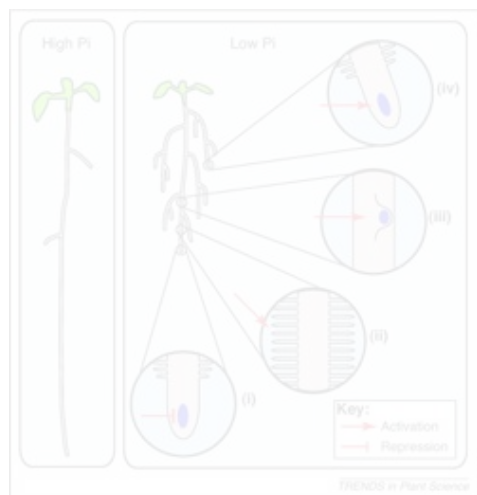
How can we solve the puzzle?

Developmental
responses



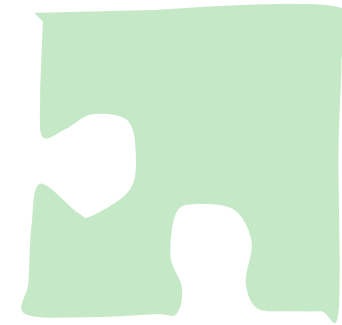
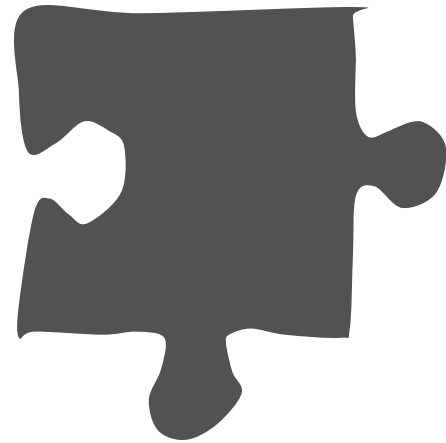
Nutritional
changes

Phosphate
transport, recovery
and recycling



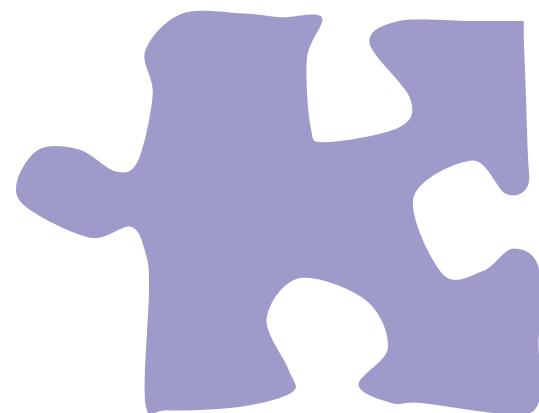
Phosphate puzzle to be solved

Regulators of
mycorrhizal
symbiosis

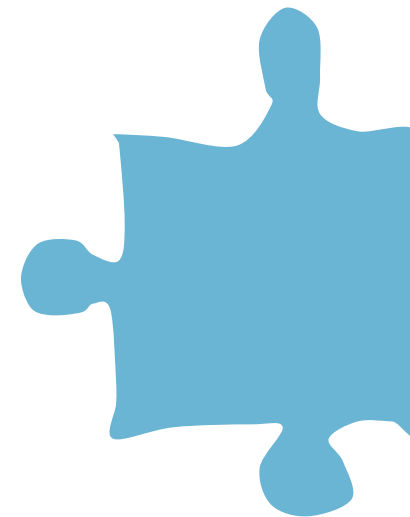


Multiple plant
species
approach

Developmental
responses



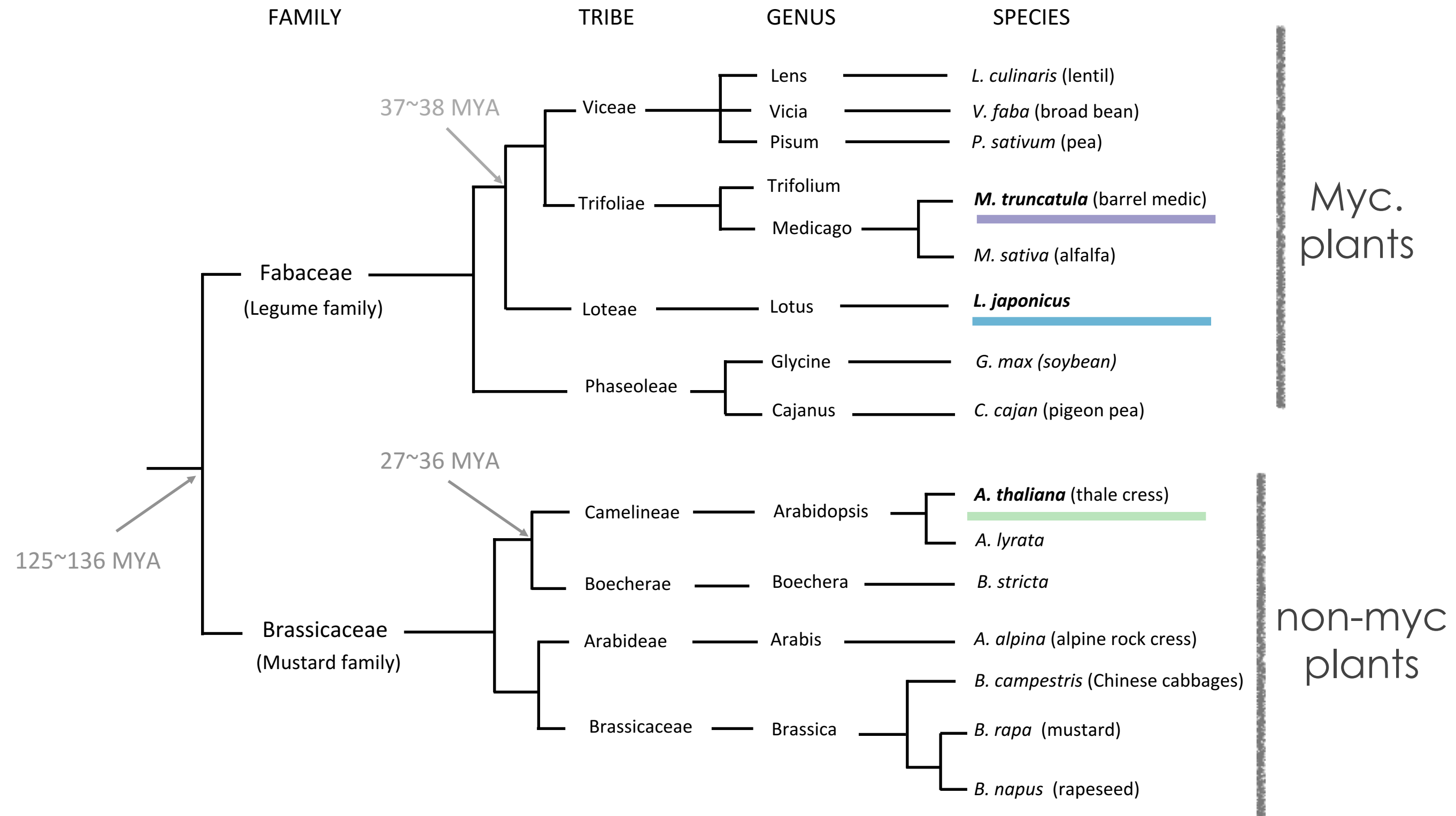
High Throughput
Root Phenotyping



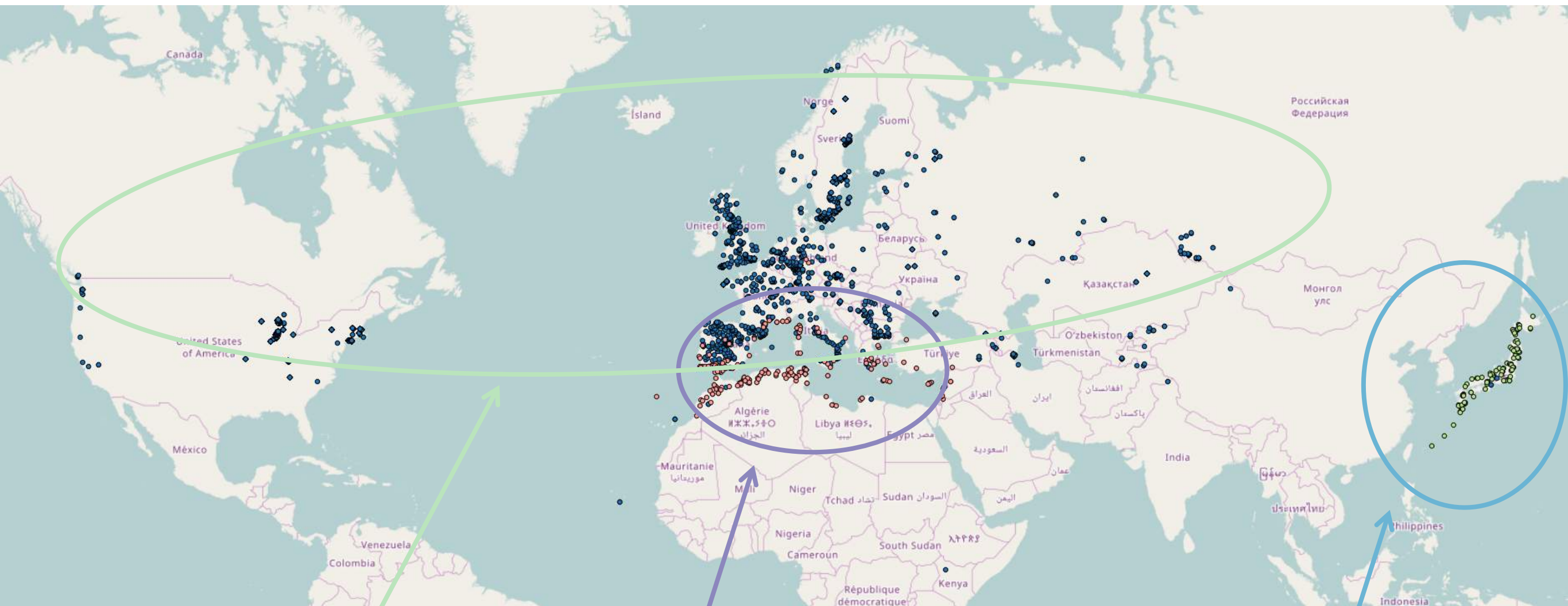
Nutritional
changes

Tissue Phosphate
quantification

Multiple plant species



Multiple plant genotypes



231 *A. thaliana*

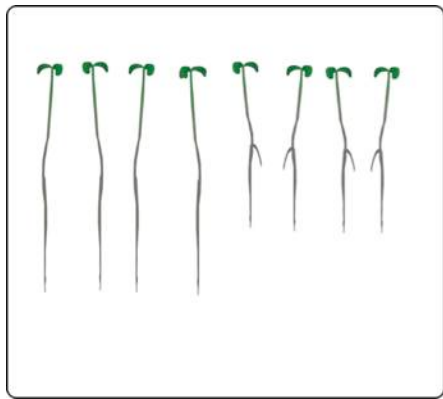
140 *M. truncatula*

130 *L. japonicus*

accession = genotype

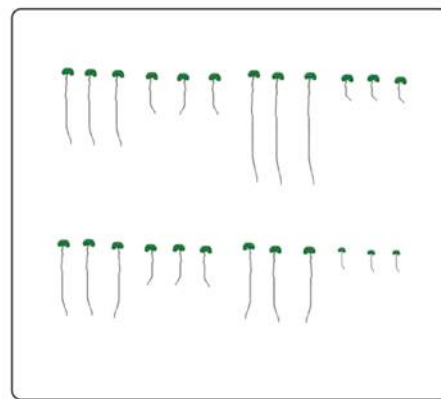
Multiple phosphate-related traits

- Automatic **root trait quantification** over low (20 μM) and high (750 μM) phosphate condition of natural populations (\rightarrow 16 traits, 2 conditions, each day)



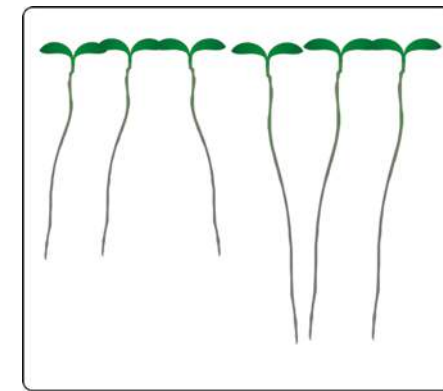
130 *Lotus japonicus*

VS.



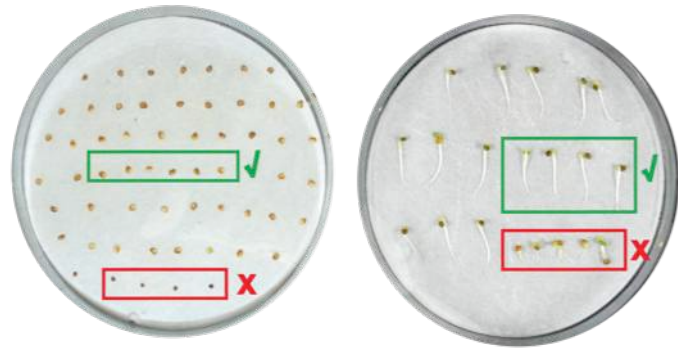
230 *Arabidopsis thaliana*
(from Santosh Satbhai)

VS.

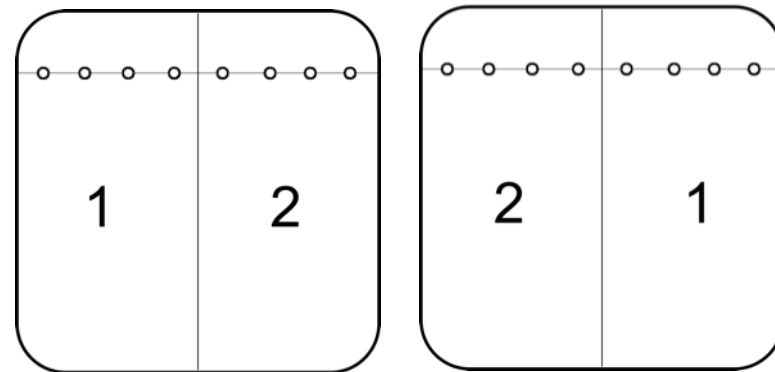


140 *Medicago truncatula*

High-throughput root phenotyping



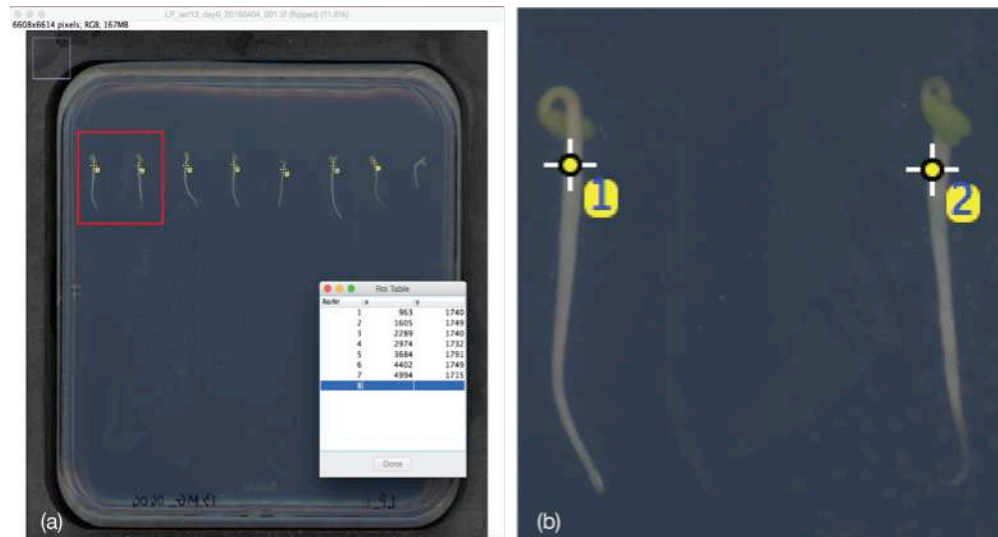
1. pre-germination



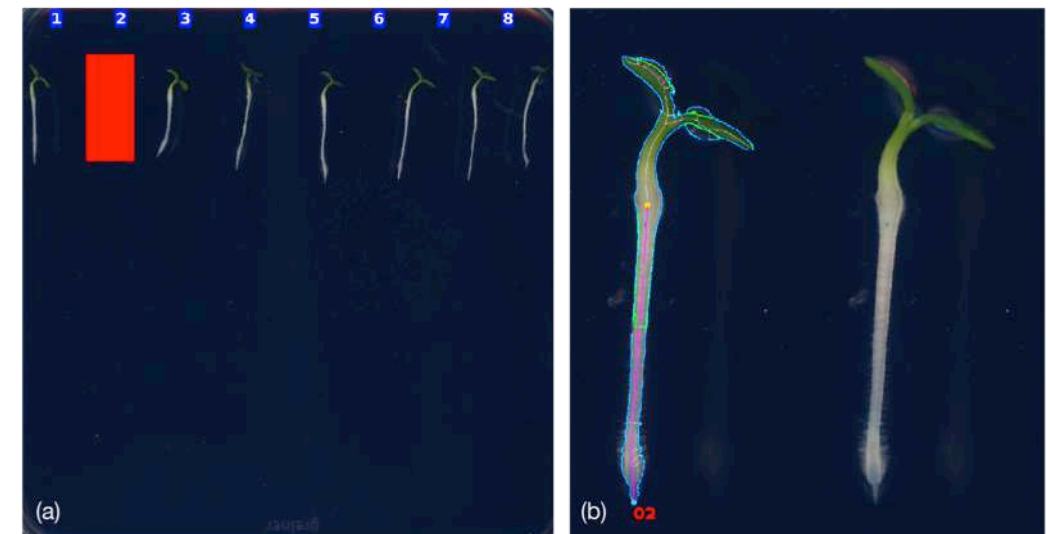
2. plant growth (8 reps)



3. HT image acquisition

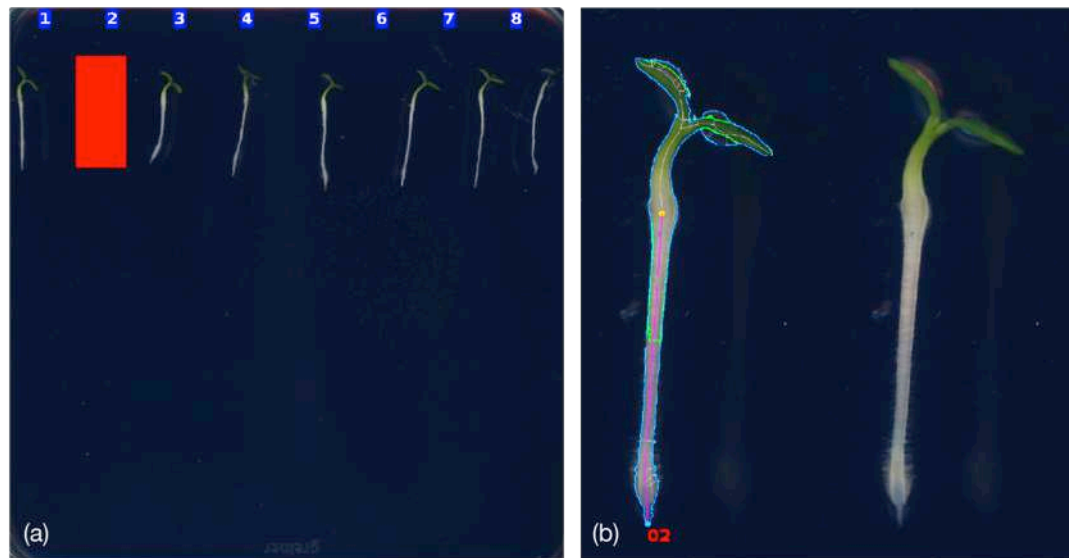


4. root identification by user



5. root segmentation

Each root gives 16 traits each day



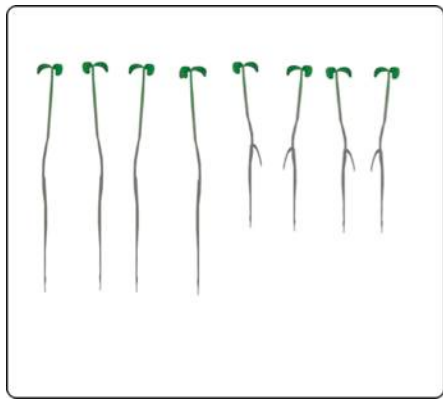
Root segmentation

Total length
Euclidian length
Root tortuosity
Root growth rate
Relative root growth rate
Root angle
Root direction index
Root horizontal index
Root vertical index
Root linearity
Average root width
Root width 20
Root width 40
Root width 60
Root width 80
Root width 100

9x
(days)

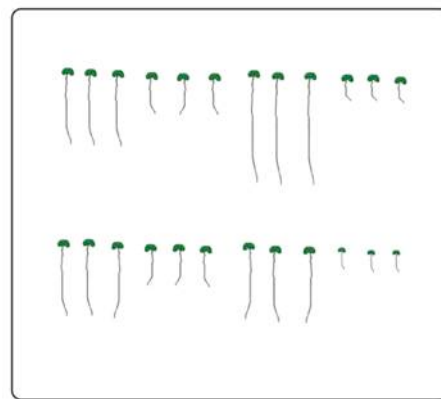
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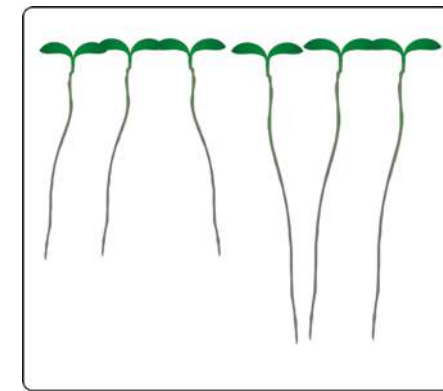
130 *Lotus japonicus*

VS.



230 *Arabidopsis thaliana*
(from Santosh Satbhai)

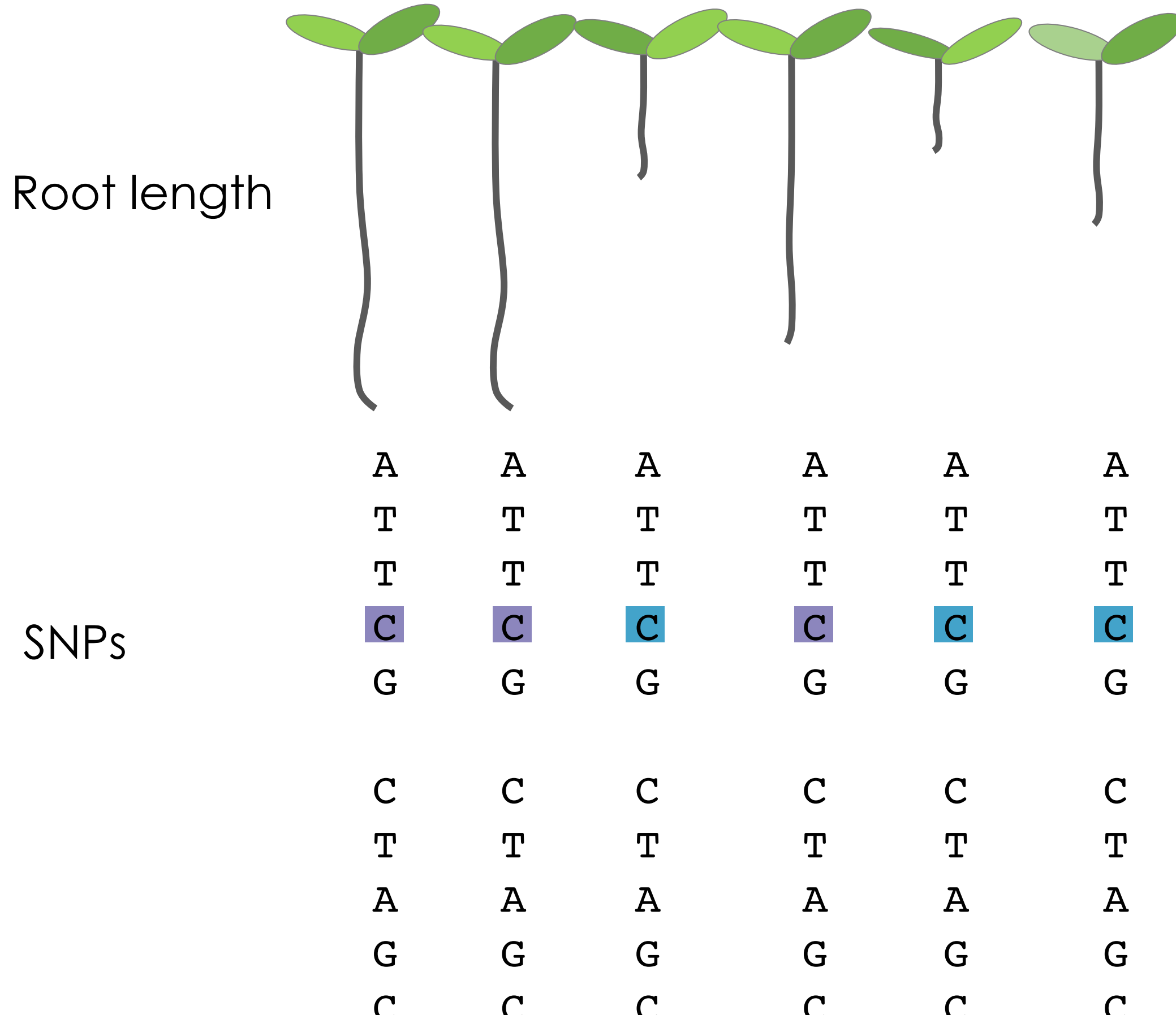
VS.



140 *Medicago truncatula*

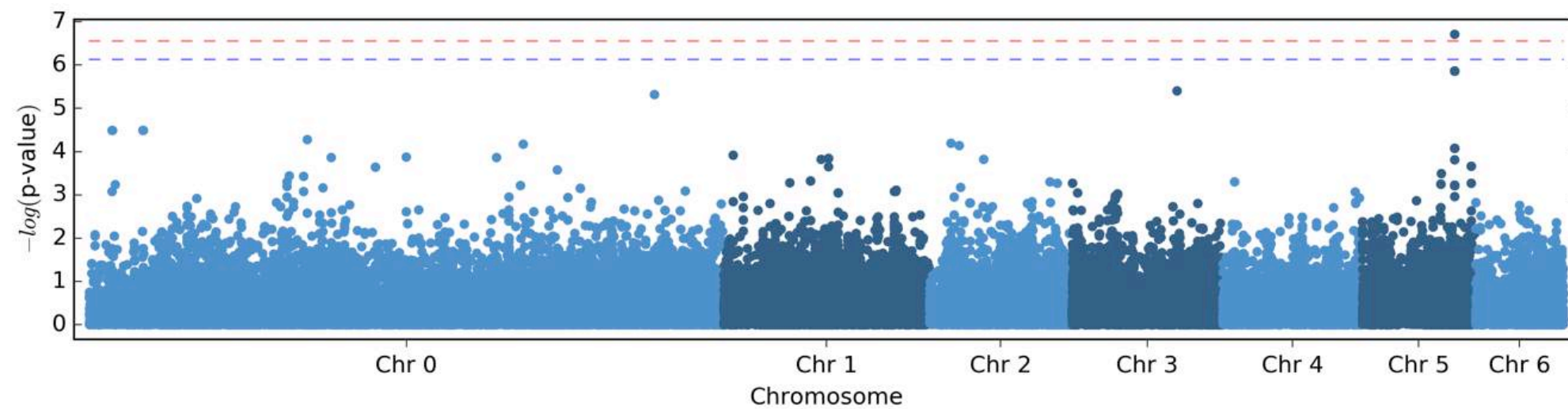
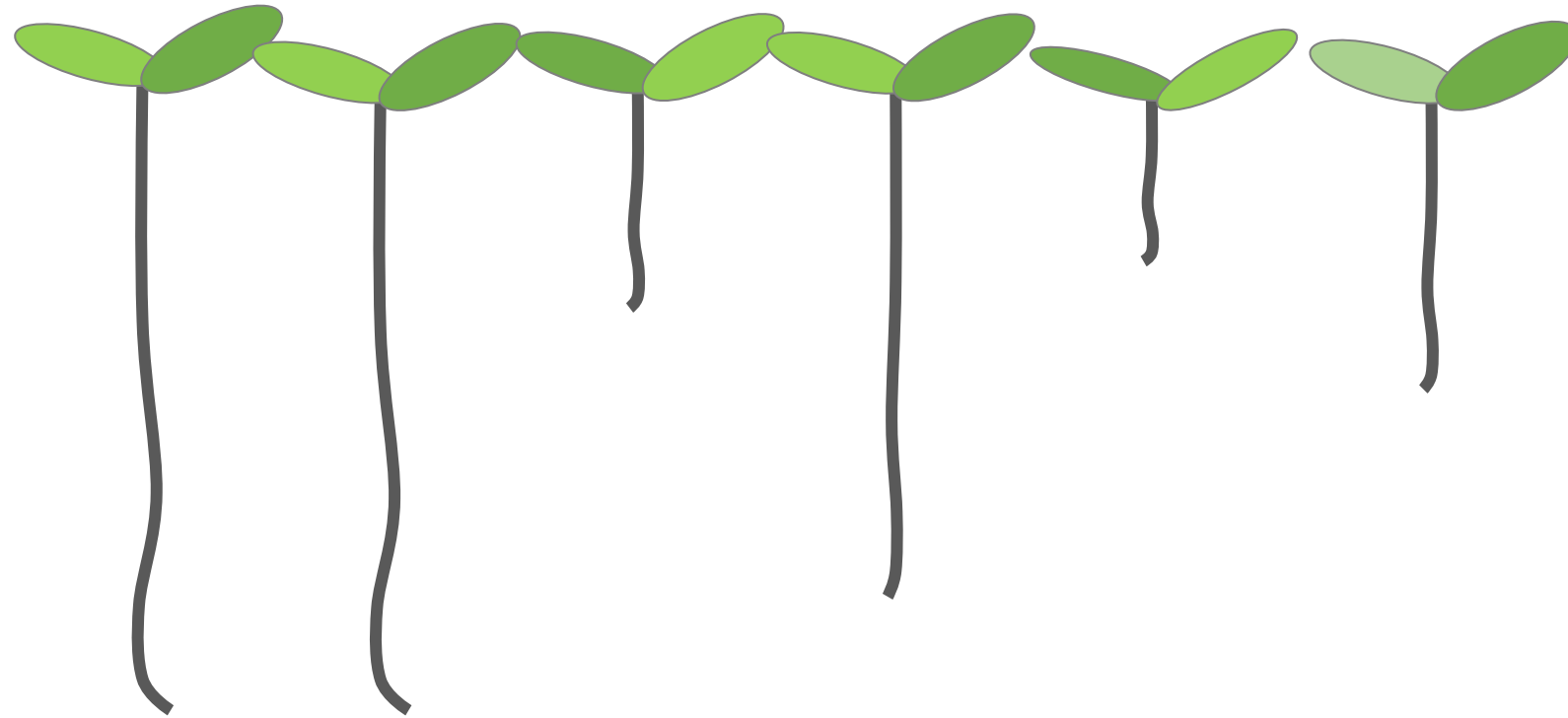
- Root and shoot **phosphate quantification** (ion chromatography) in 130 Lotus accessions

Genome Wide Association Study (GWAS)

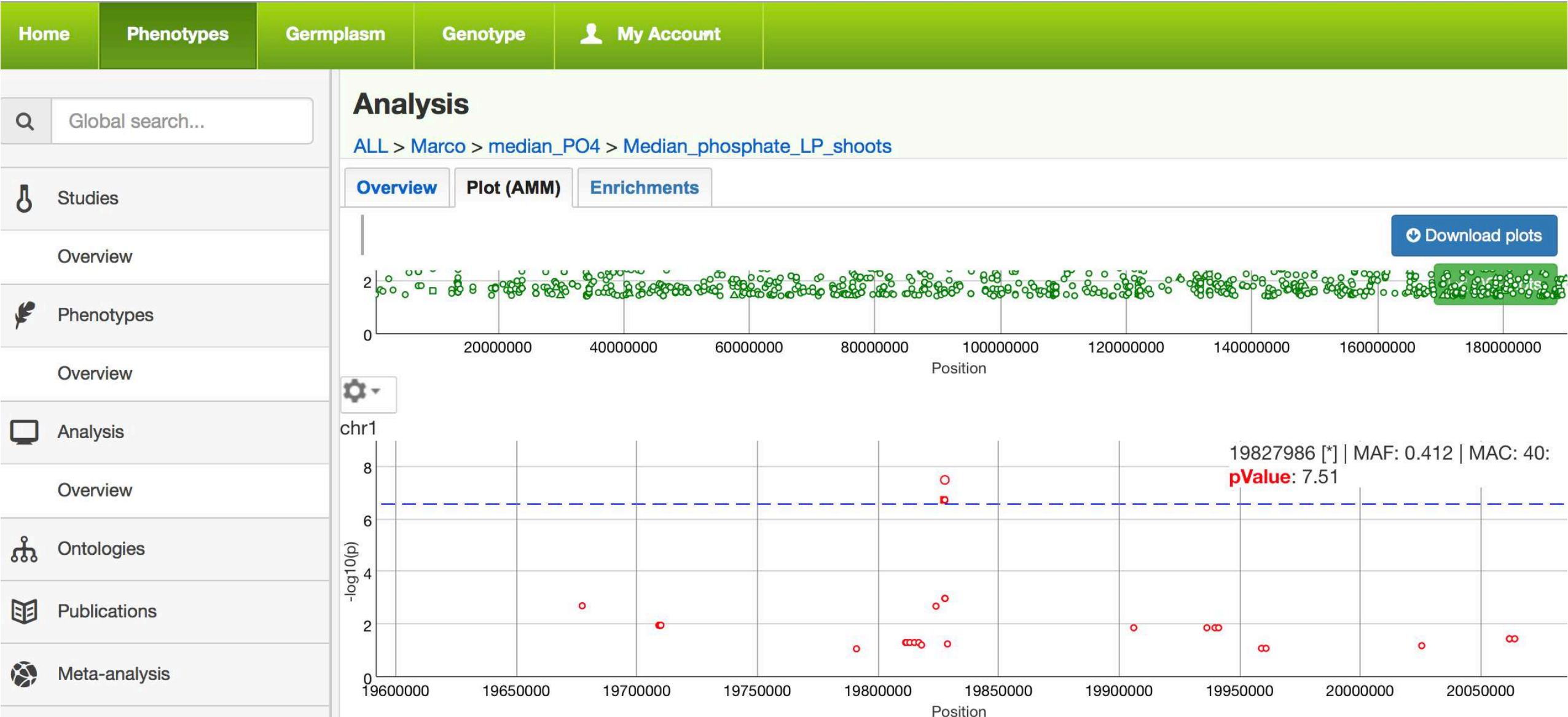


Genome Wide Association Study (GWAS)

Root length



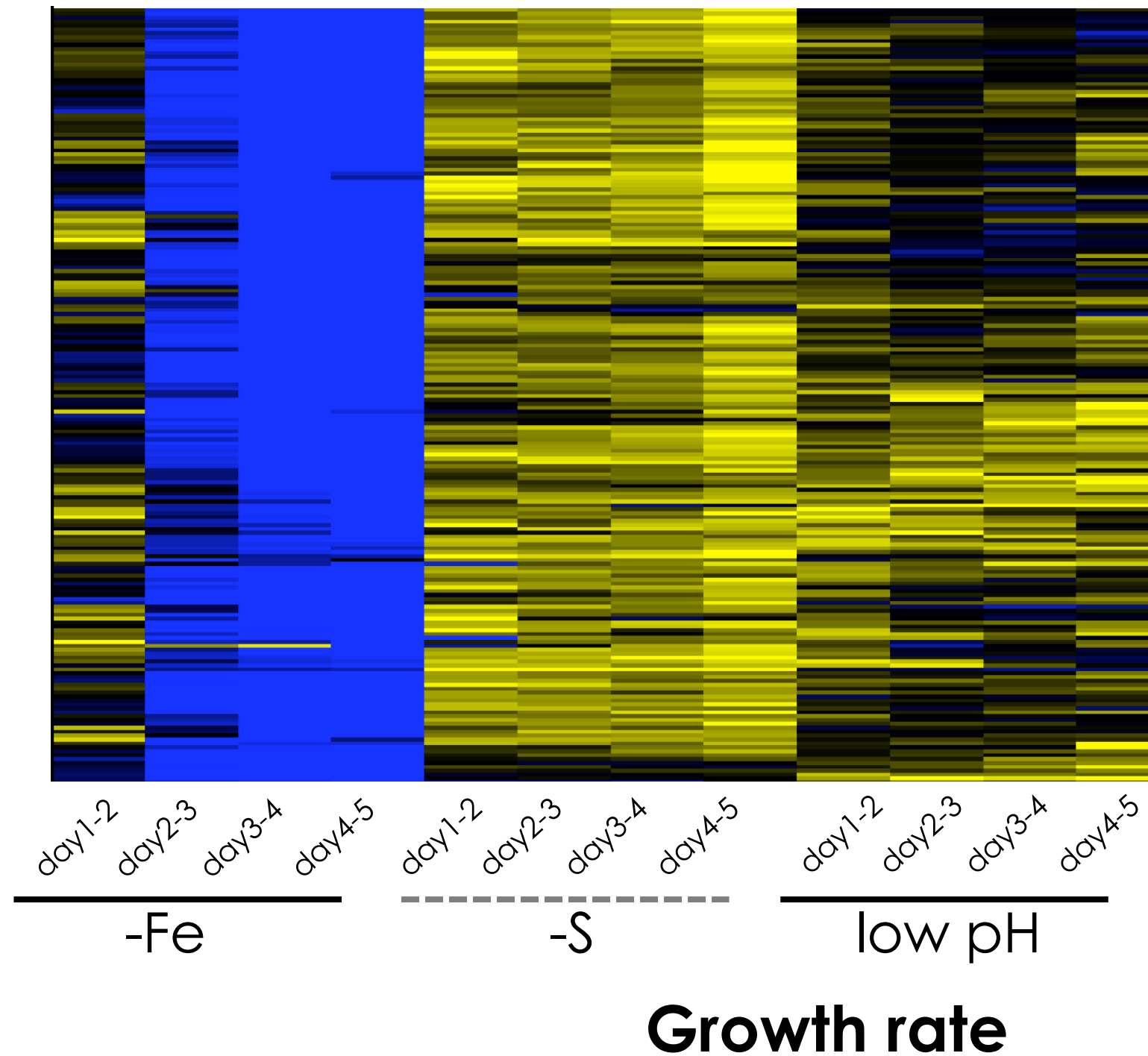
GWAPP: A web application for GWAS in Arabidopsis and Lotus (lotus.au.dk/gwas)



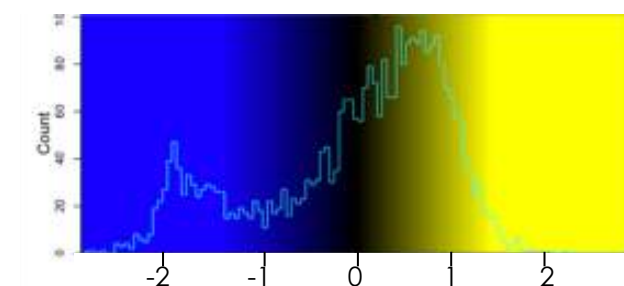
Aims

1. Are mycorrhiza forming plant species responding to phosphate deficiency in a manner similar to *Arabidopsis thaliana* ?
2. Are root growth traits correlated to nutrient accumulation capacity?
3. Are variants of orthologous genes associated with P-related traits in different species?

Low phosphate has a unique bimodular effect on *Arabidopsis* root growth



231 *A. thaliana* accessions



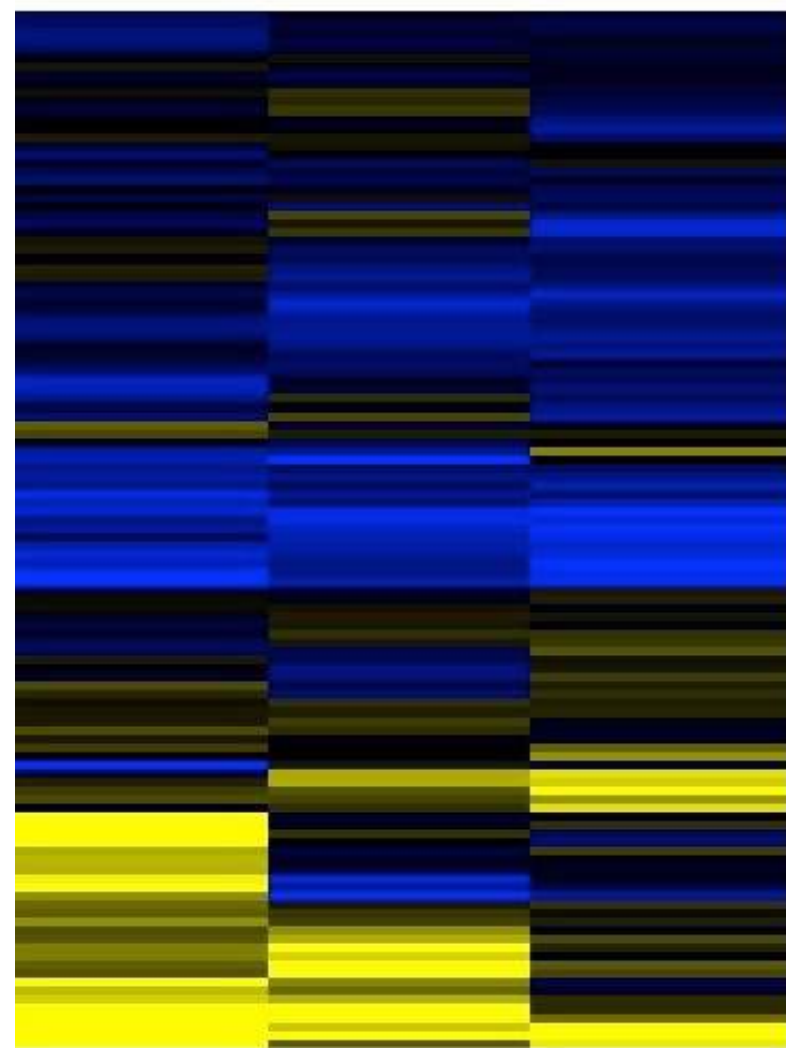
data from Santosh Satbhai



Both *Medicago* and *Lotus* have a genotype-dependent bimodal responses to low phosphate



130 *Lotus* accessions



day6-7

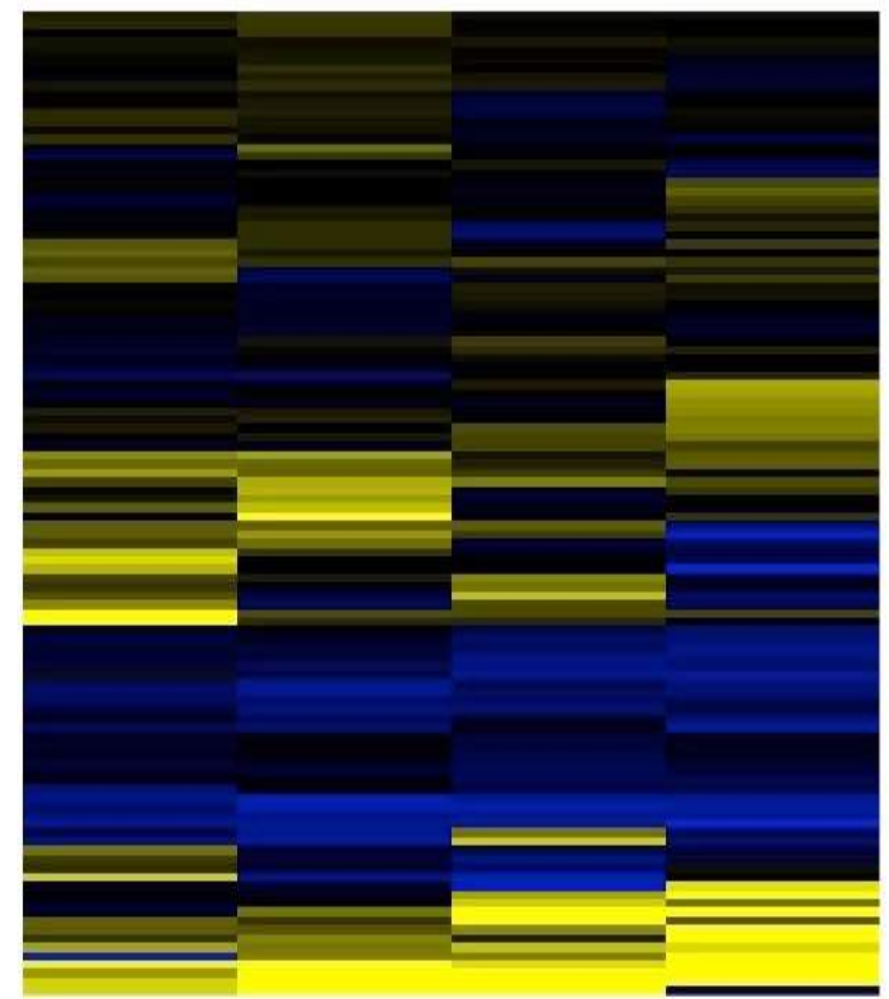
day7-8

day8-9

Growth rate



140 *Medicago* accessions



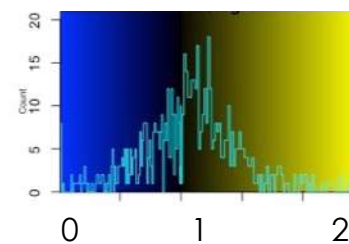
day1-2

day2-3

day3-4

day4-5

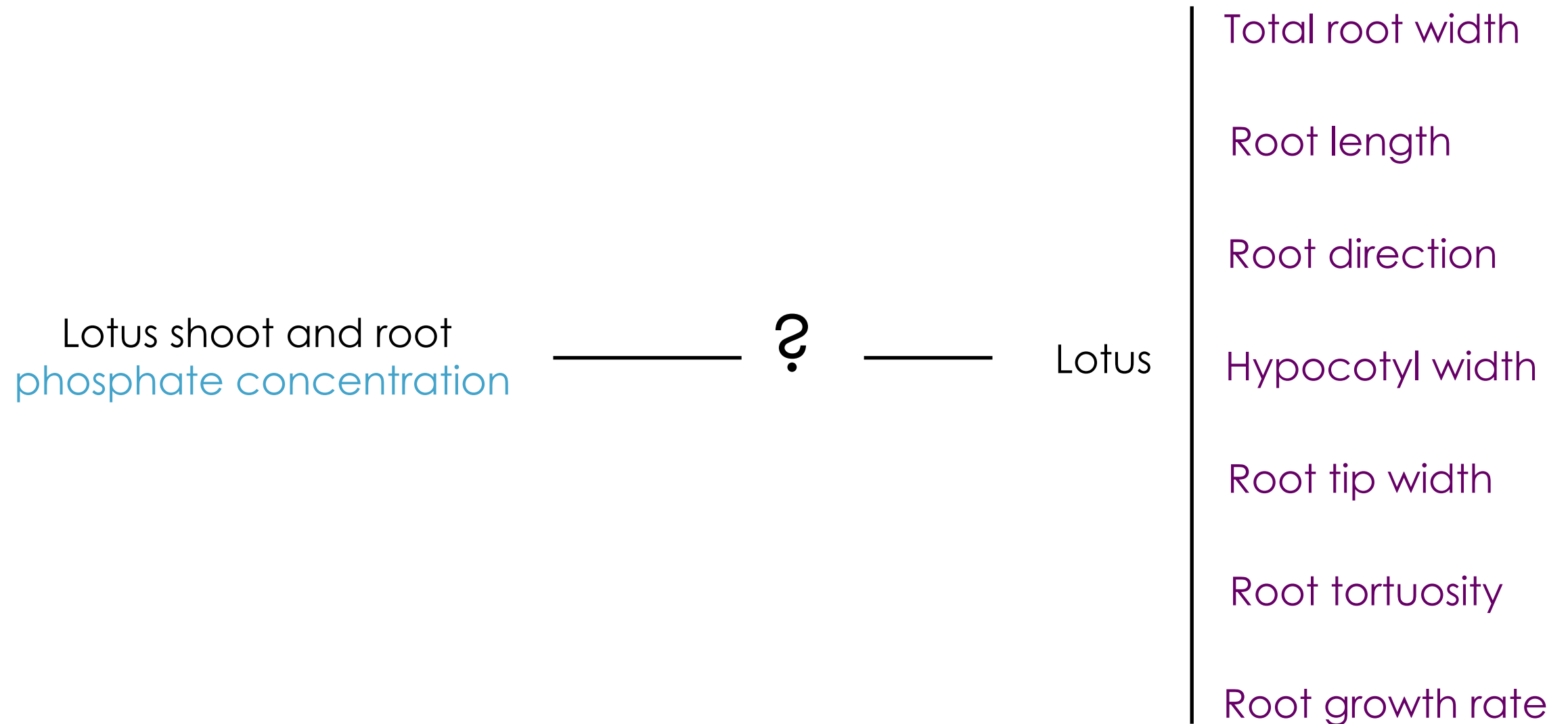
Growth rate



Aims

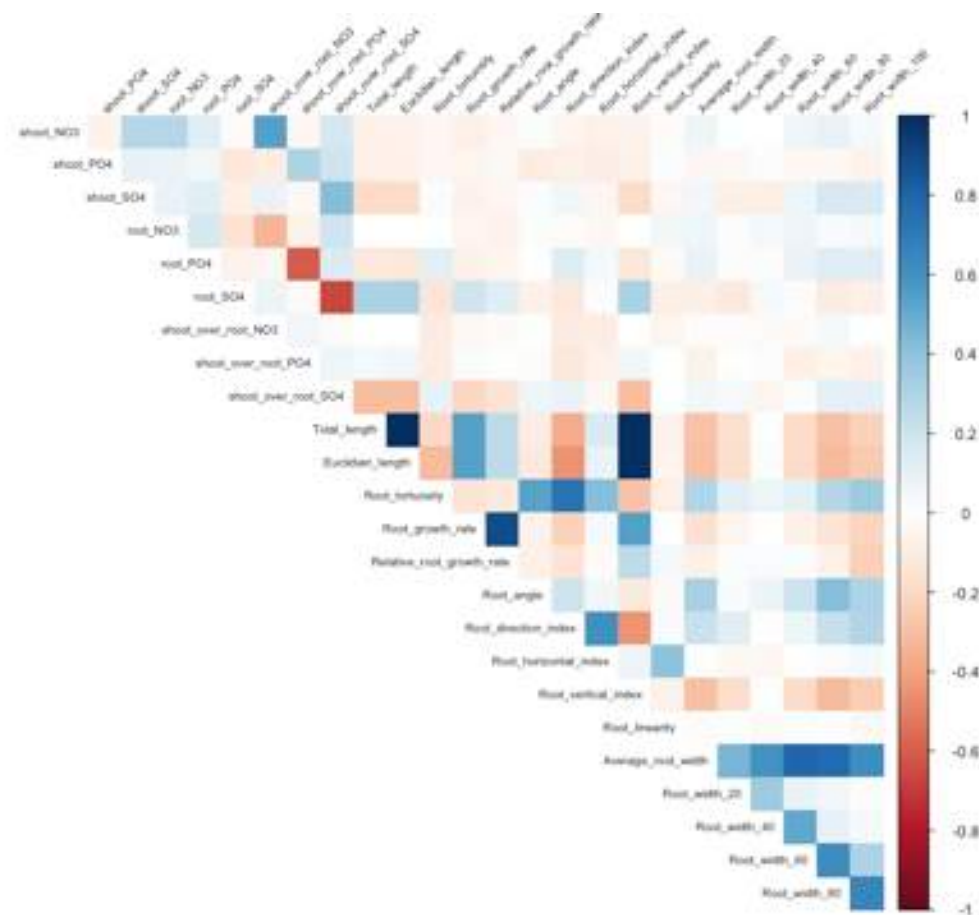
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2. Are root growth traits correlated to nutrient accumulation capacity?

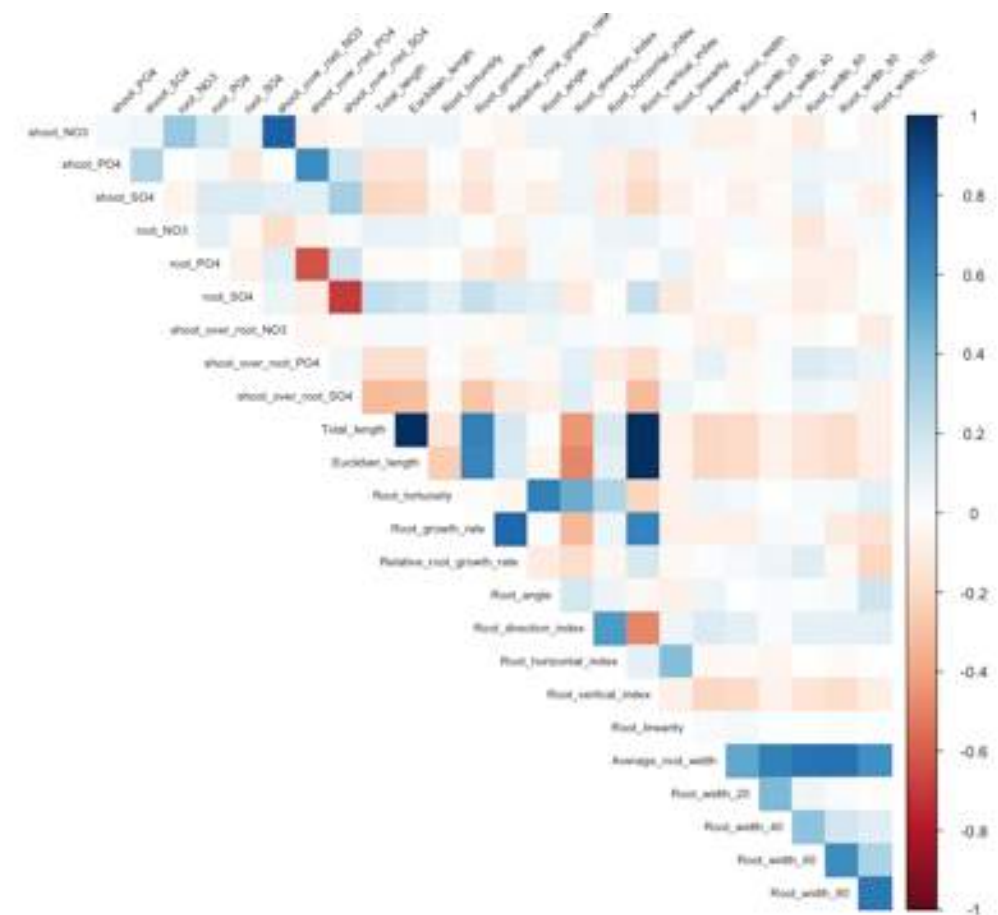


In Lotus most of correlations among traits do not depend on P levels

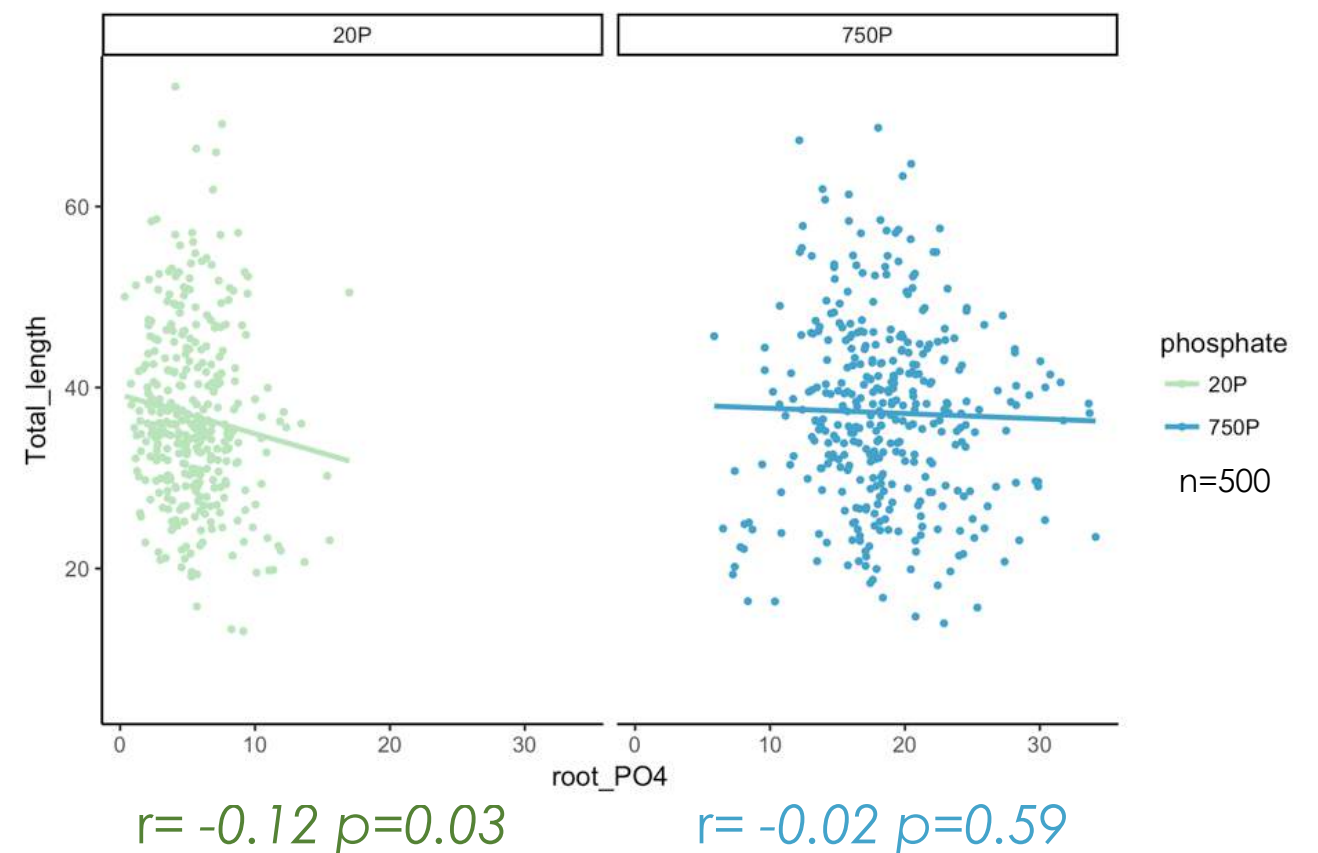
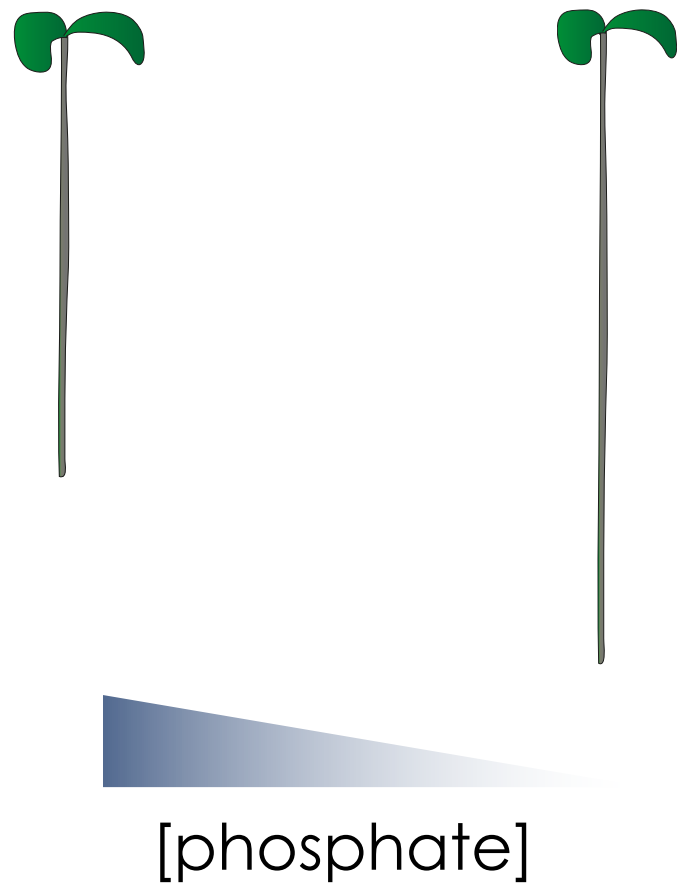
Traits over LP



Traits over HP

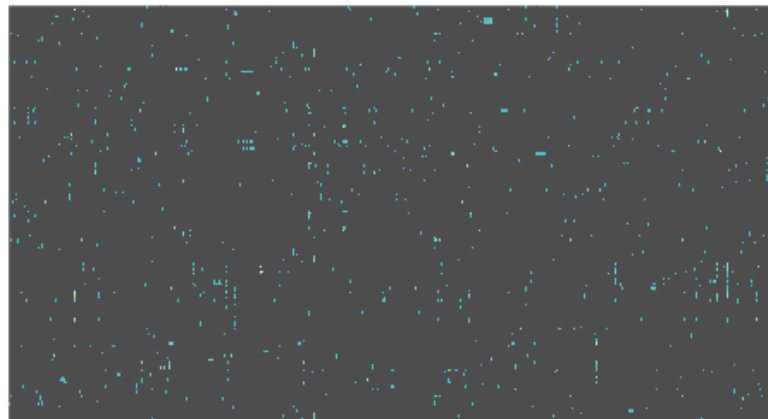


Only in **low phosphate** medium, root length negatively correlates with phosphate concentration



Overlapping Lotus GWAS from different traits pinpoint to a single locus

Lotus
root system
architecture
traits



Genomic position

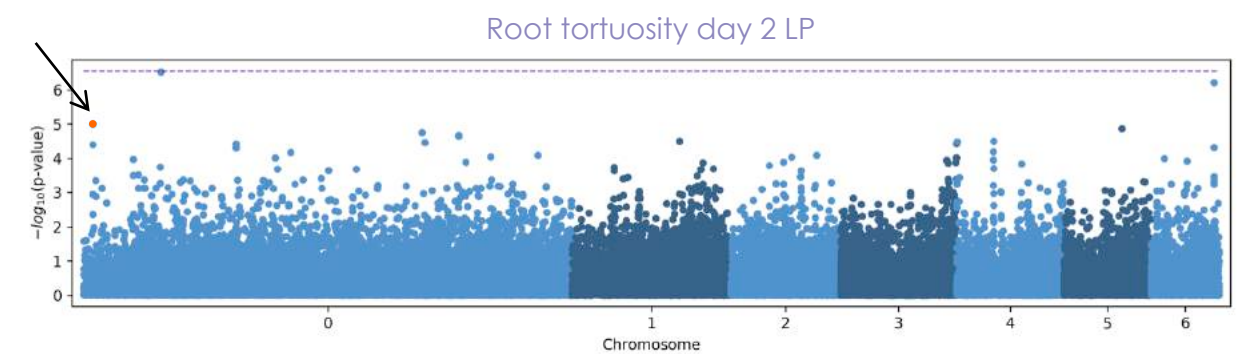
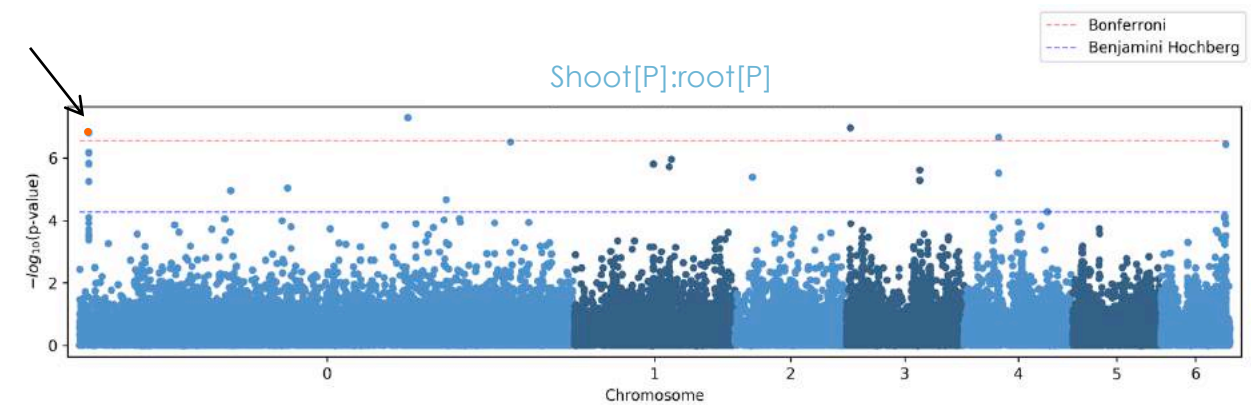
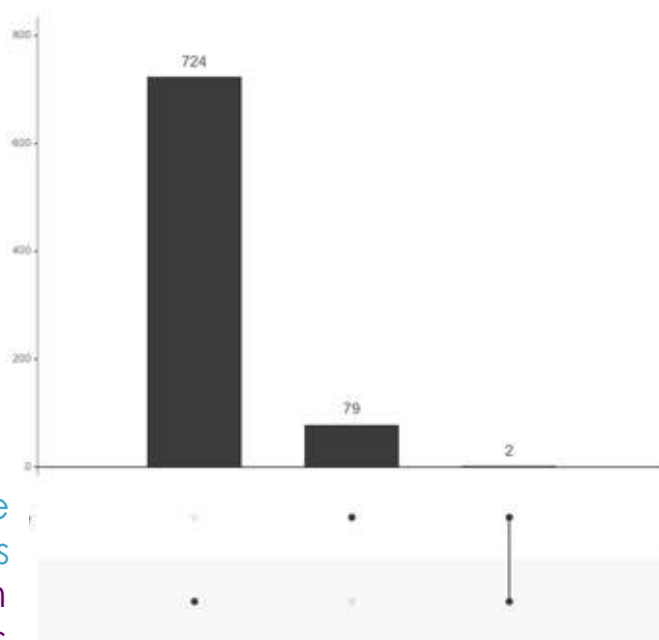
VS.

Lotus
phosphate
accumulation
traits

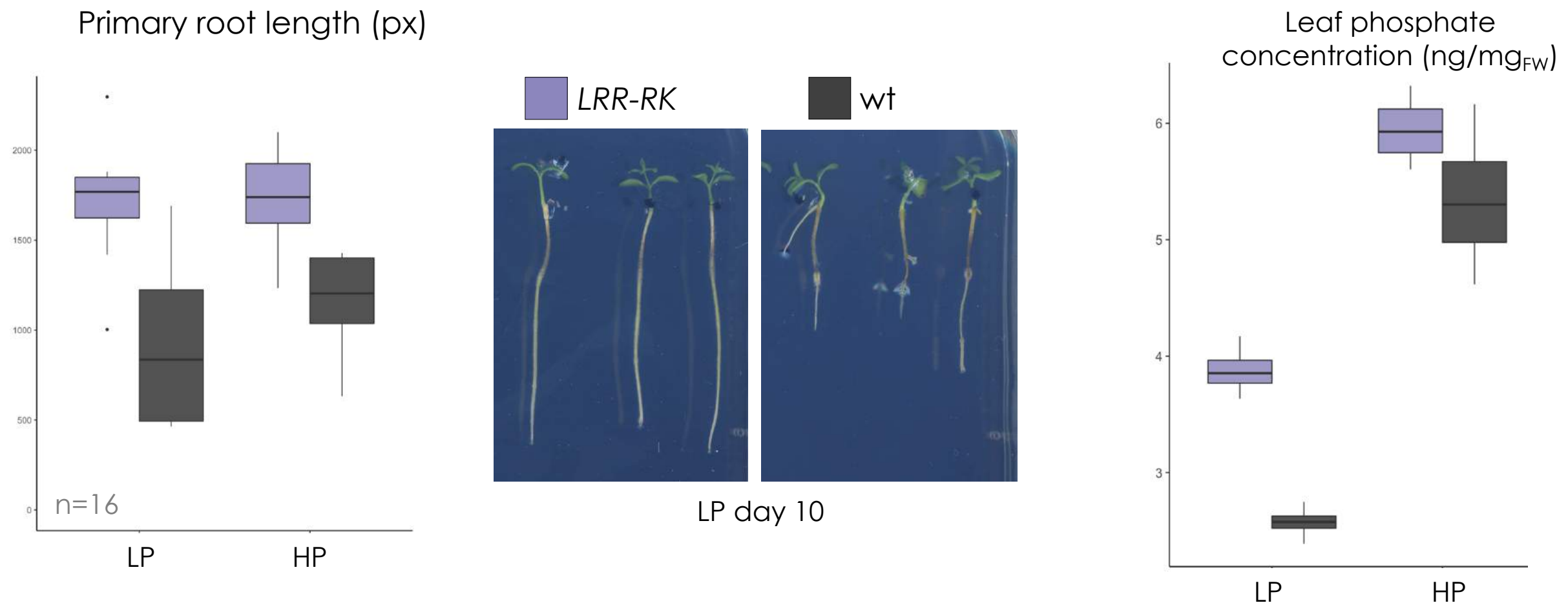


Genomic position

Phosphate
accumulation traits
Root system
architecture traits



A cell surface receptor, LRR-RK, associated with different P-dependent traits, controls primary **root growth** and **phosphate accumulation** in Lotus leaves



- A possible receptor of root to shoot phosphate signaling?
- Low level of expression (hard to spot with other approaches)



Aims

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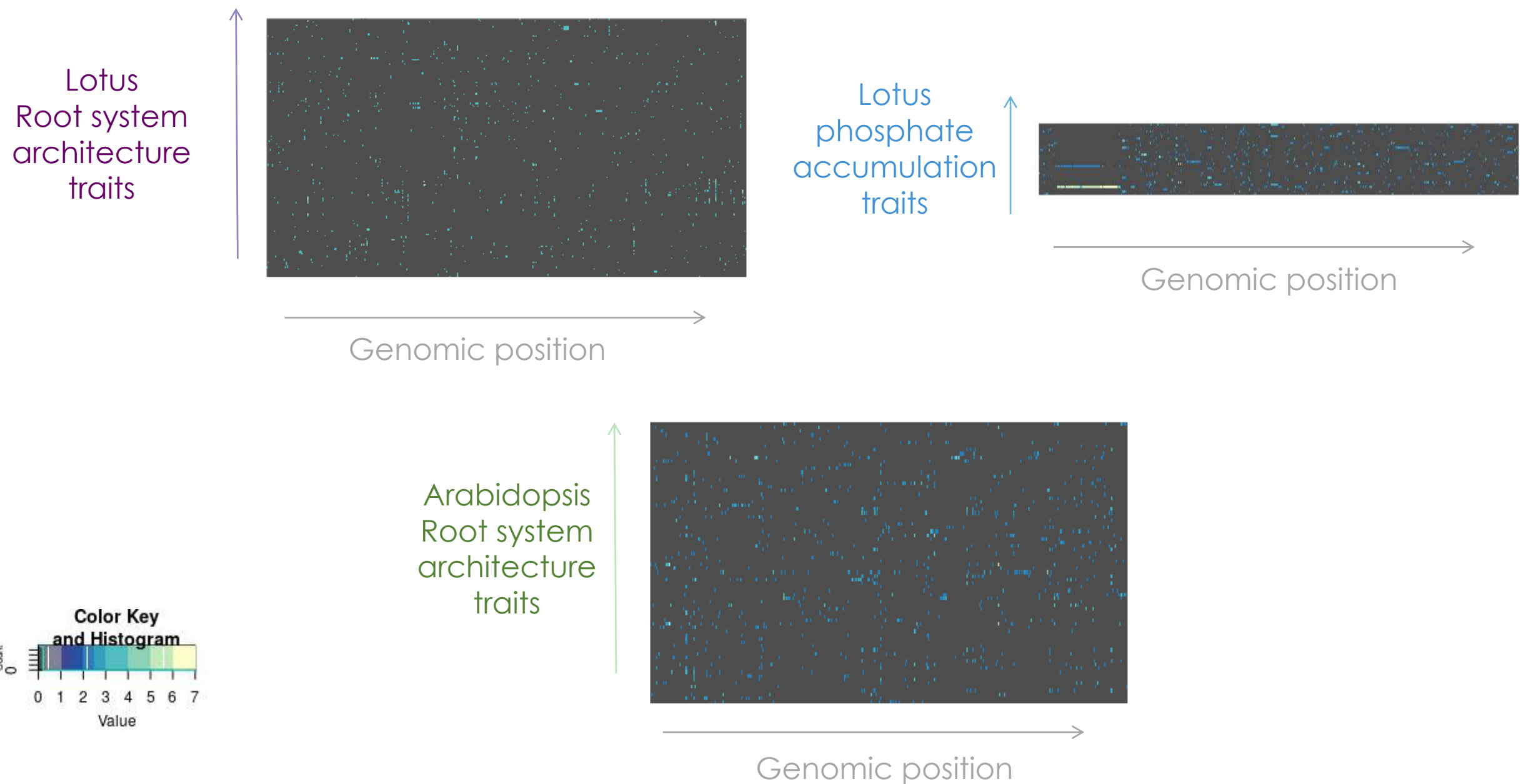
High Throughput
Root Phenotyping

Multi plant
species
approach

Tissue Phosphate
quantification



Overlapping GWAS from different plant species pinpoints to a few loci



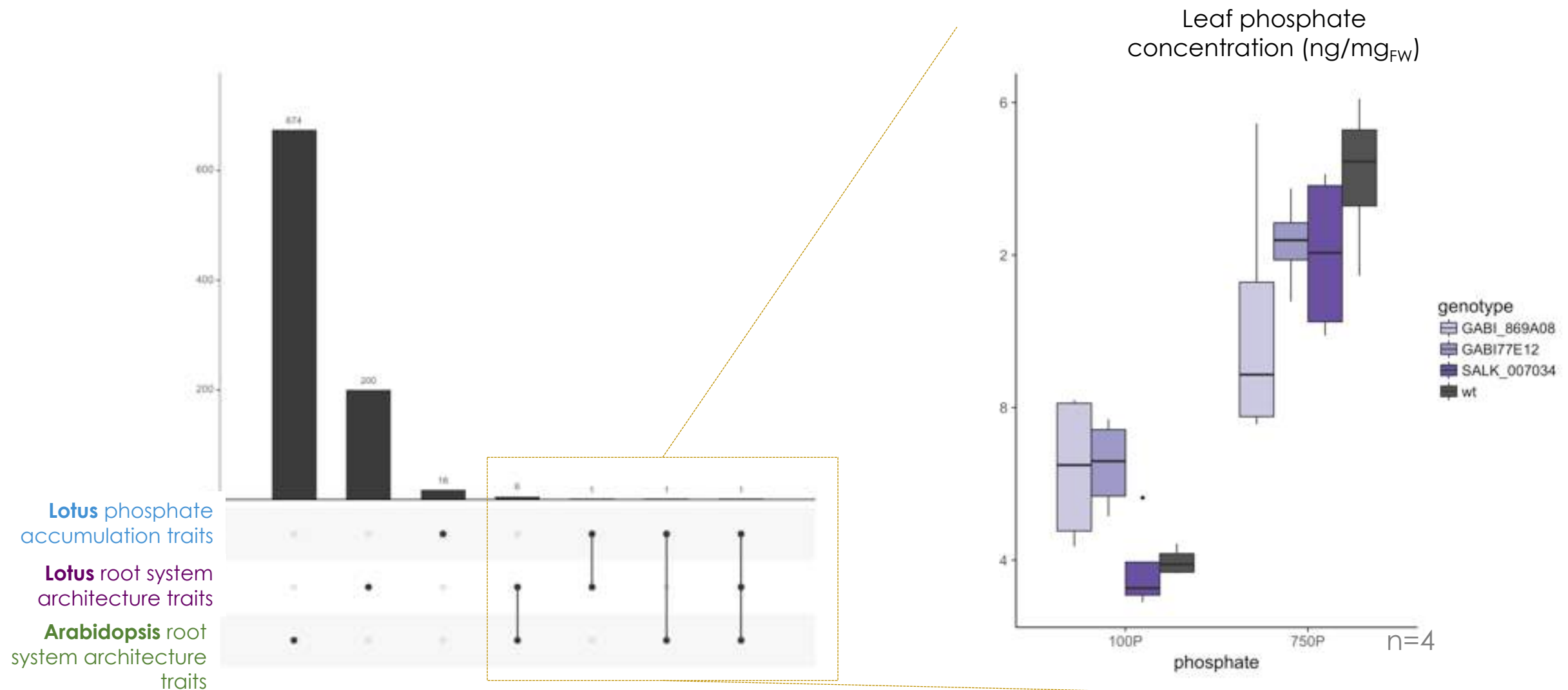
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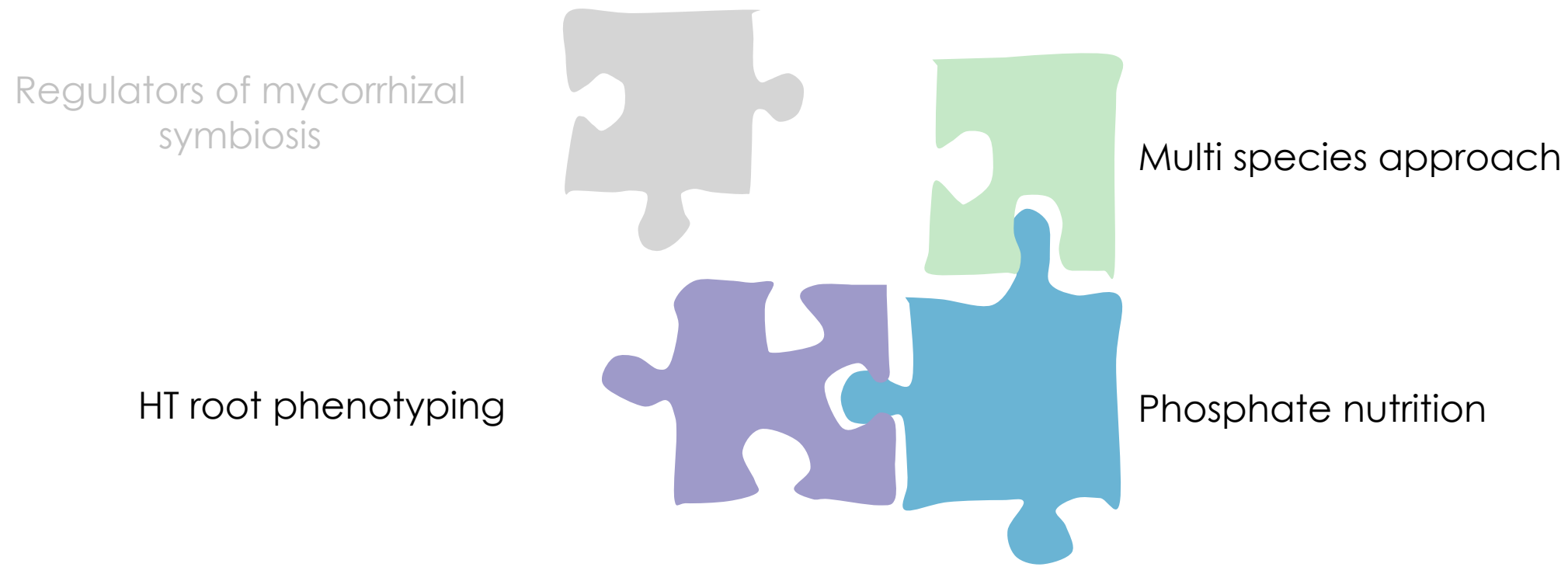


Variation in orthologous genes in Arabidopsis and Lotus is associated with P responses less than expected by chance



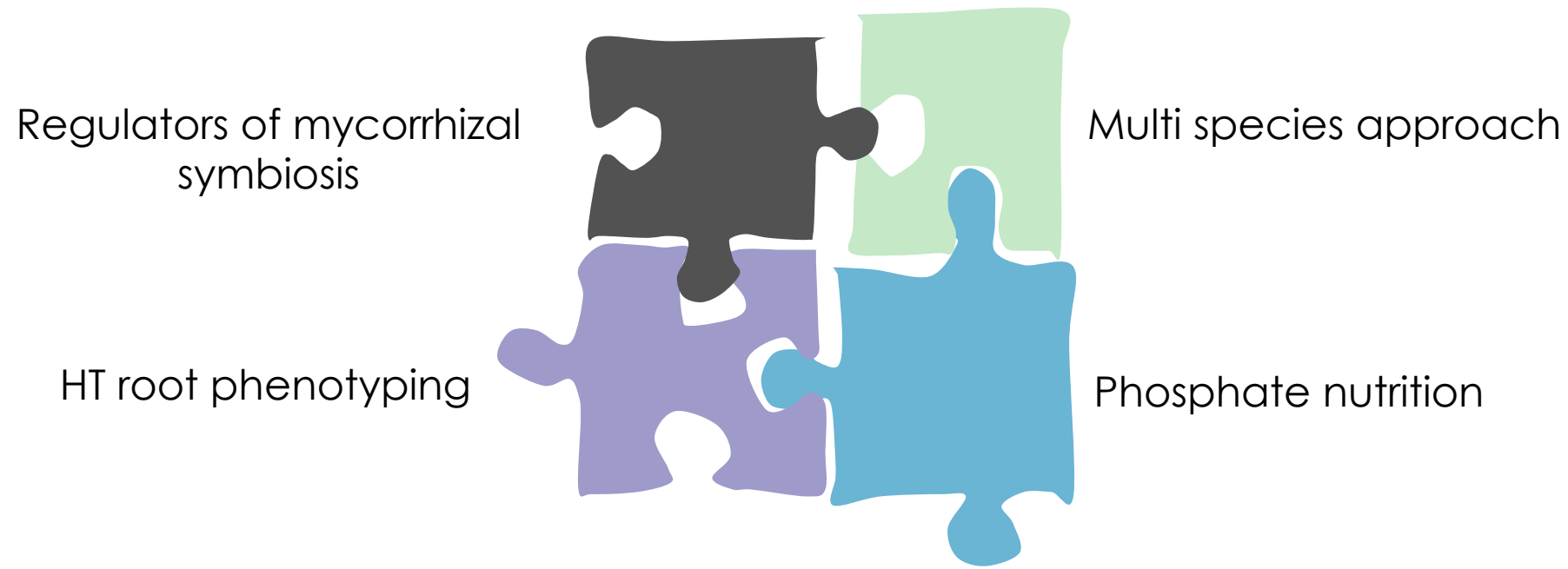
- The number of overlaps among plant species is less than what expected by chance (10000 permutation test on the same gene lists)
- 3 out of 3 genes selected are responsible for different phosphate accumulation in Arabidopsis (based on available data for 300 mutants, $p=0.02$ to get it by chance).
- 1 mutant line available in Lotus shows the same phenotype

Conclusions



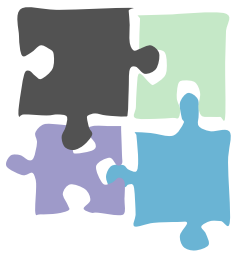
- Lotus is a feasible plant for HT root phenotyping
- Integration of phosphate accumulation and root growth highlights true positive genes in phosphate
- A cell surface receptor, Leucine-Rich Repeat Receptor Kinase, regulates root growth and phosphate accumulation in Lotus
- Comparing GWAS hits from different plant species, we identified three genes involved in phosphate accumulation

What's next?

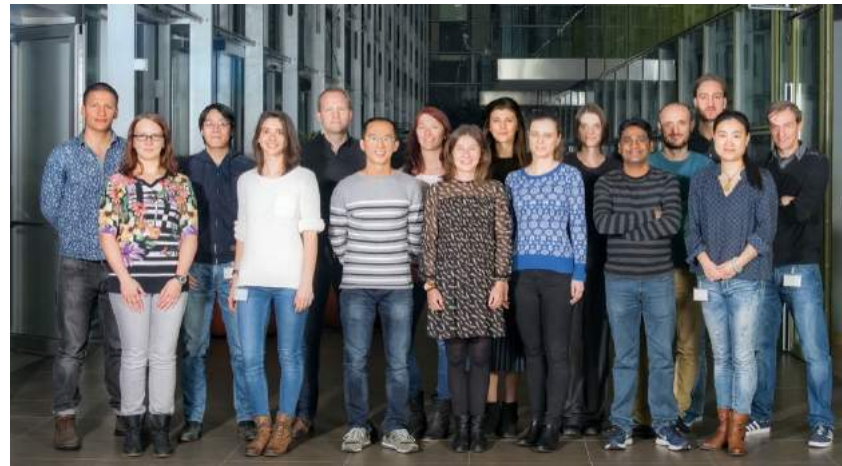


- Understand at which level the cell surface protein is regulating phosphate levels in Lotus (root to shoot signal?)
- Comparing Lotus vs. Medicago GWAS overlap (phospholipase)
- Dissecting Lotus mutants for their capacity of forming mycorrhizal interaction over different phosphate levels
- Set up a root phenotyping system for later stages of growth

Acknowledgements



Wolfgang Busch
Christian Goeschl
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Samantha Krasnodebski
and the group



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University of Cologne

Stig Andersen
University of Aarhus

Medicago
Hapmap project

Yasin Dagdas &
group



VBCF Plant
Facilities

Lab Service

Nordborg group
Uemit Seren
Rahul Pisupati



Thank you for your attention!

Lotus japonicus as a model plant

Sequenced genome (v3.0)

Self pollinating plant species

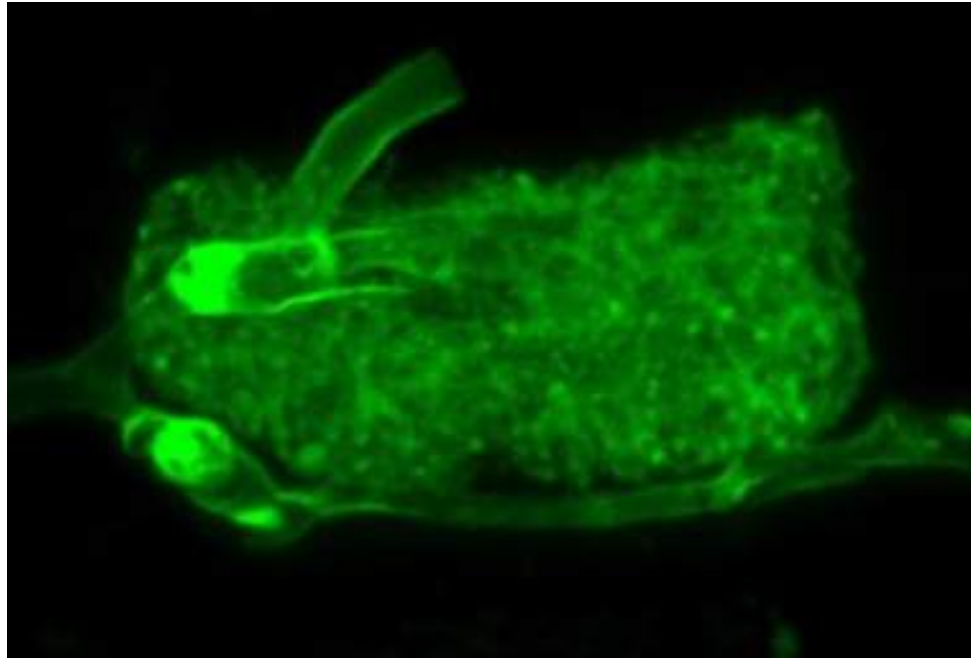
More than 100k insertional mutant lines
with 640k annotated insertions

Lotus Base (lotus.au.dk): an integrated
online platform

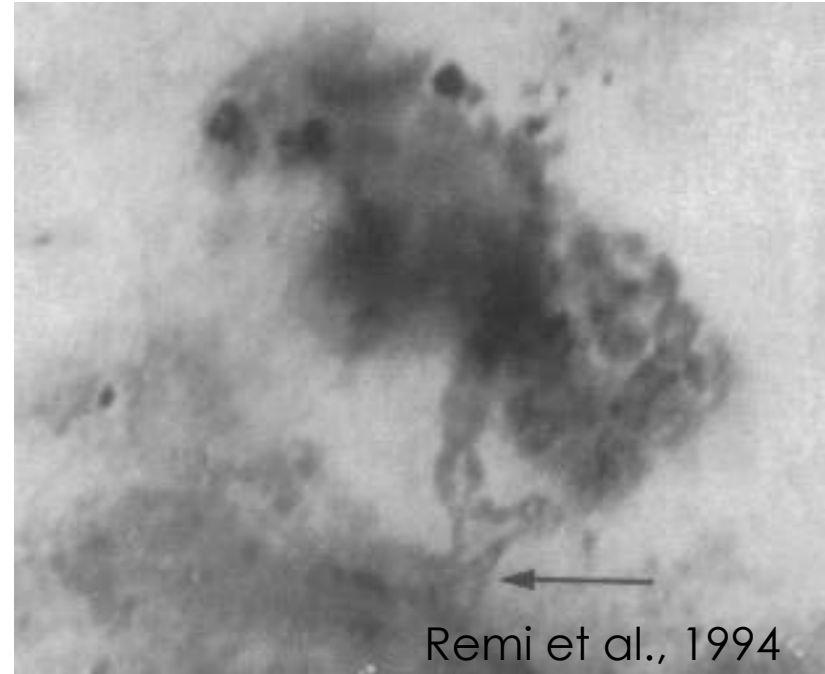
User-friendly online GWAS platform
(lotus.au.dk/gwas)



Now and then

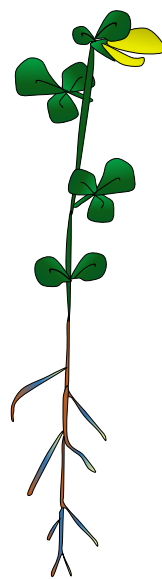


Today



Remi et al., 1994

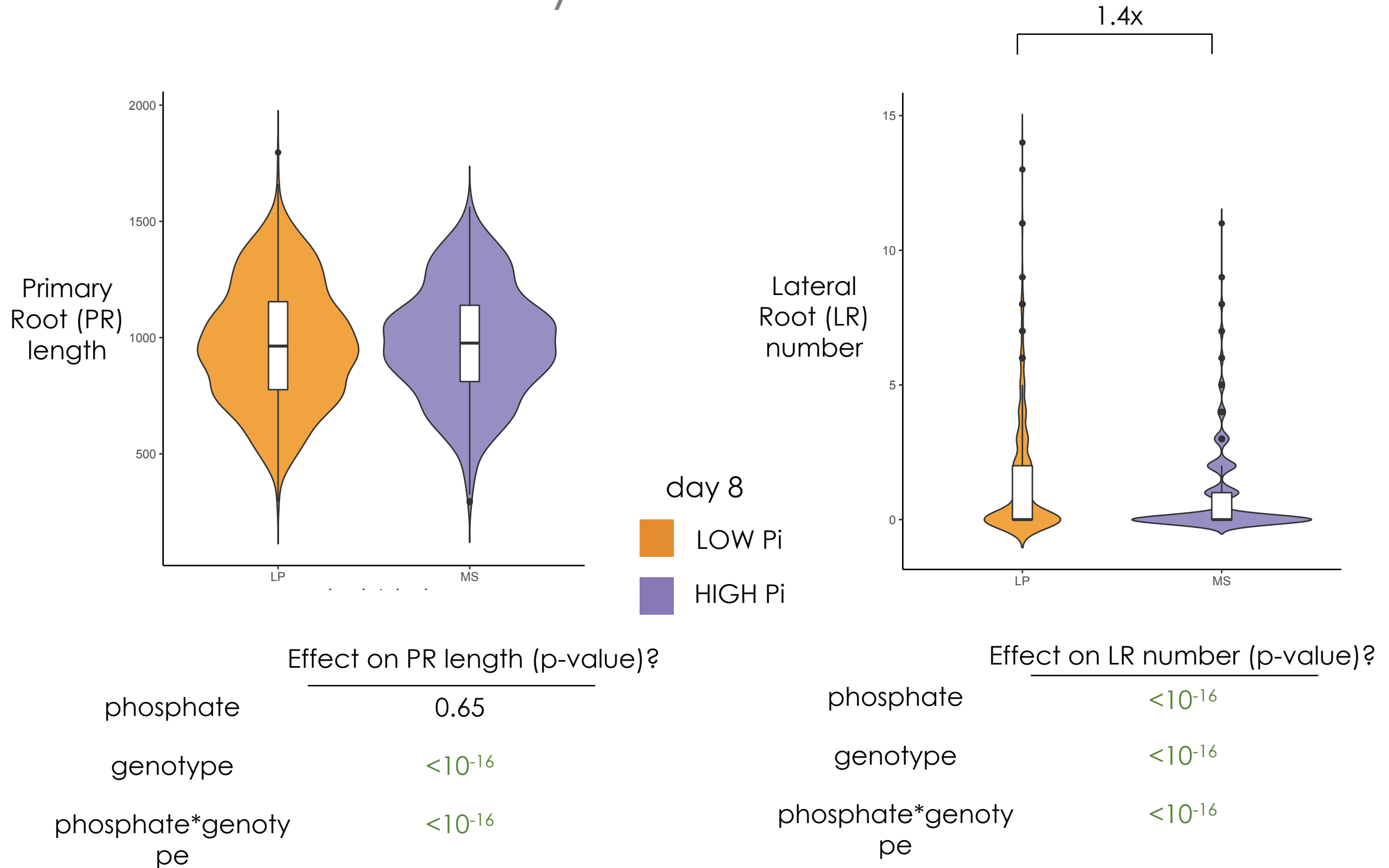
Some 400 MYA



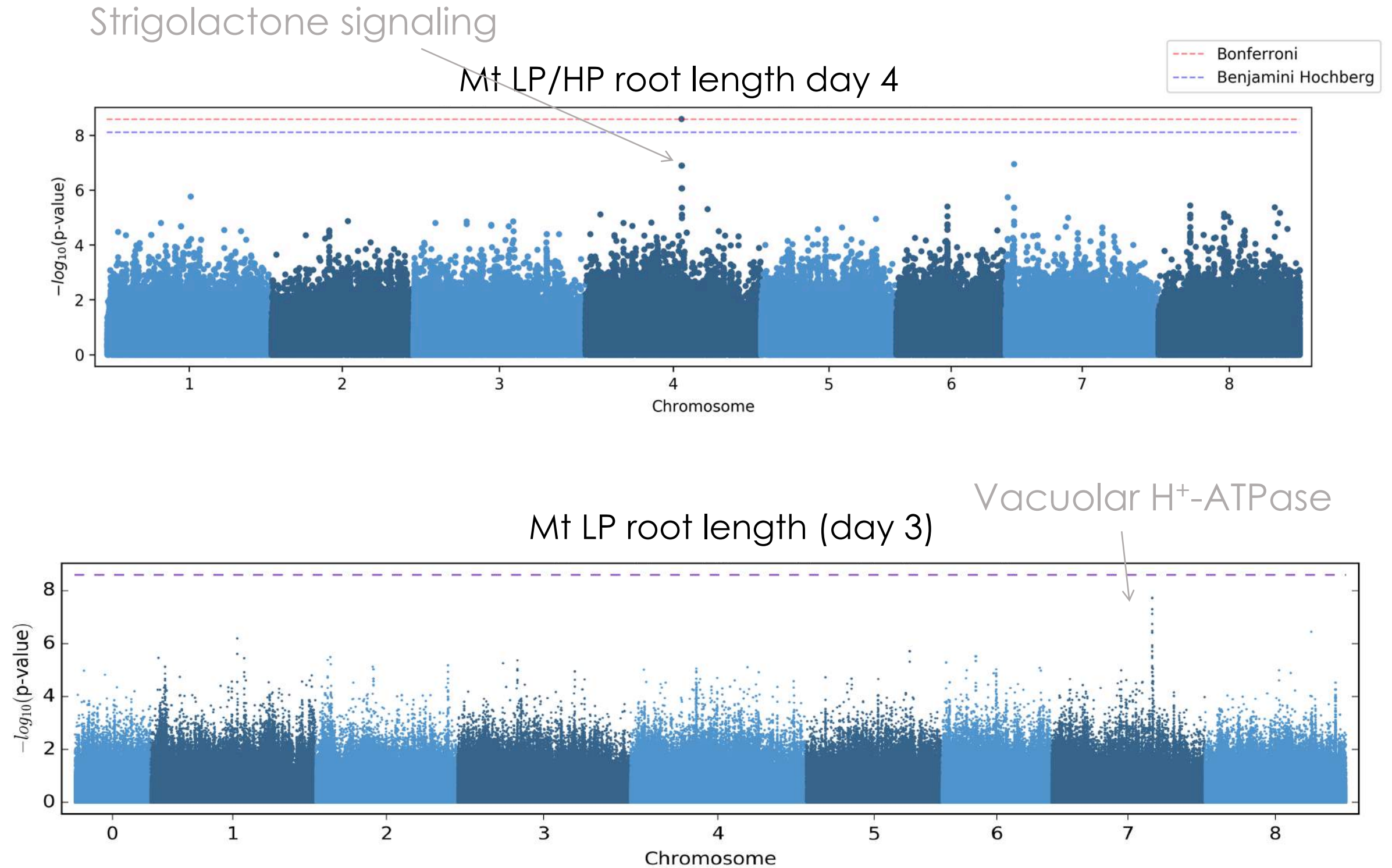
Aglaophyton major



Effects of phosphate deficiency on *Medicago truncatula* Root System Architecture

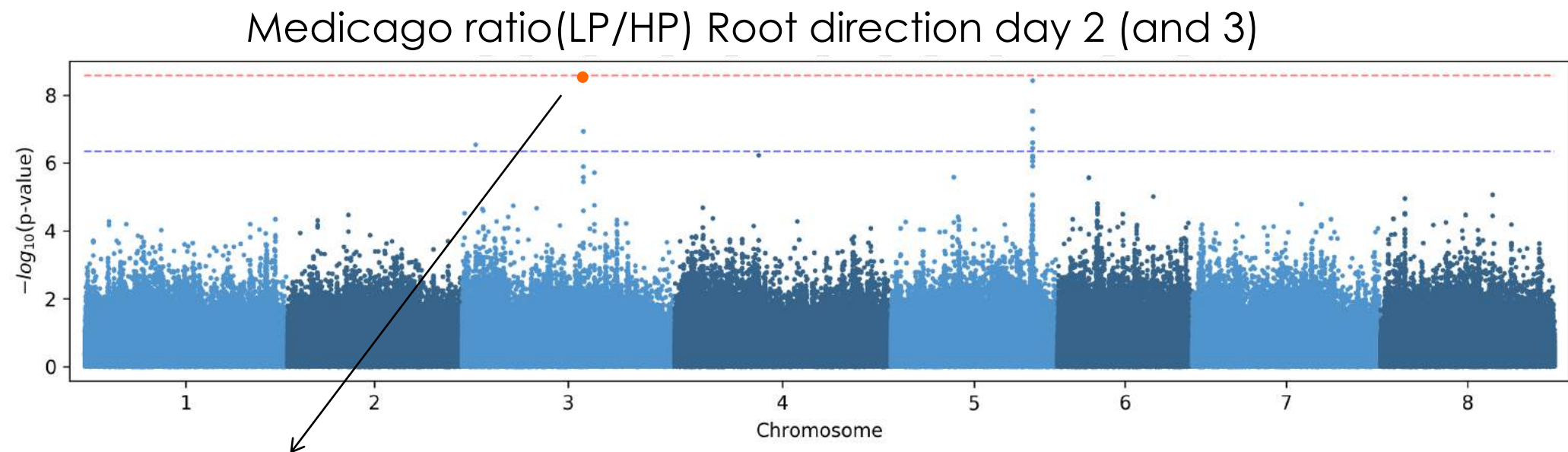


Candidate genes regulating root growth in Medicago

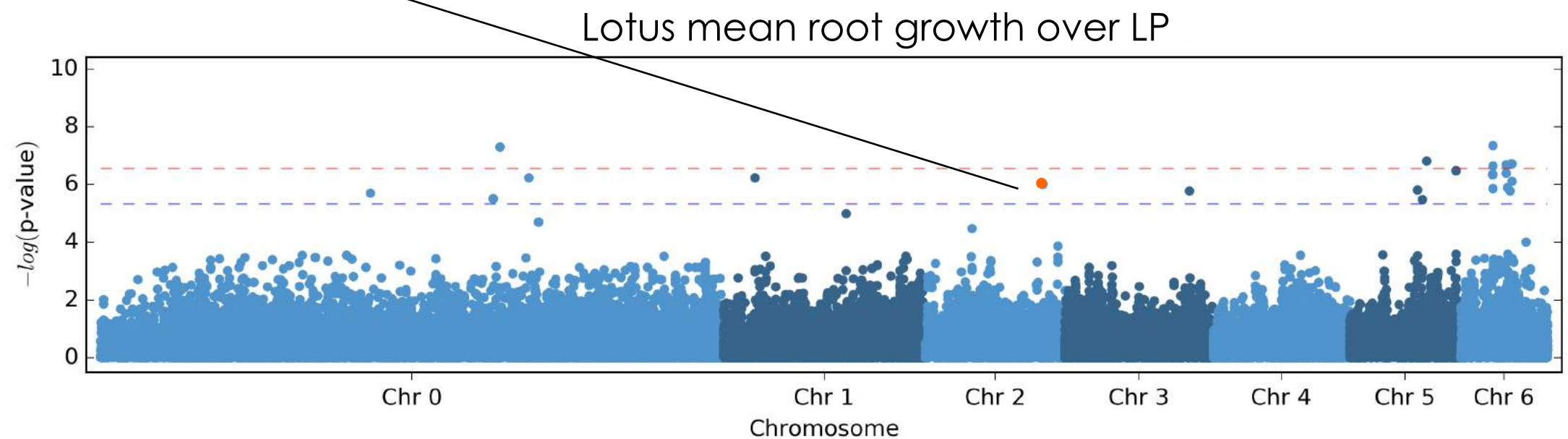


Based on an imputation of Medicago genome (40M SNPs) with Beagle

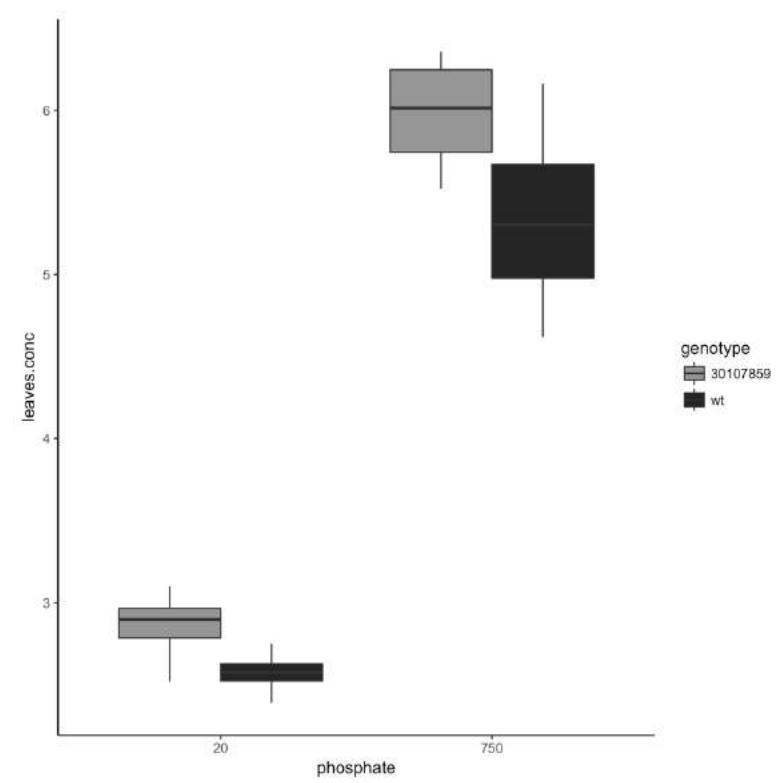
Orthologue loci in Lotus and Medicago are both associated with variation in root responses to phosphate



phosphoinositide-specific phospholipase C



Both involved in lipid changes upon phosphate stress?
Or lipid signaling...

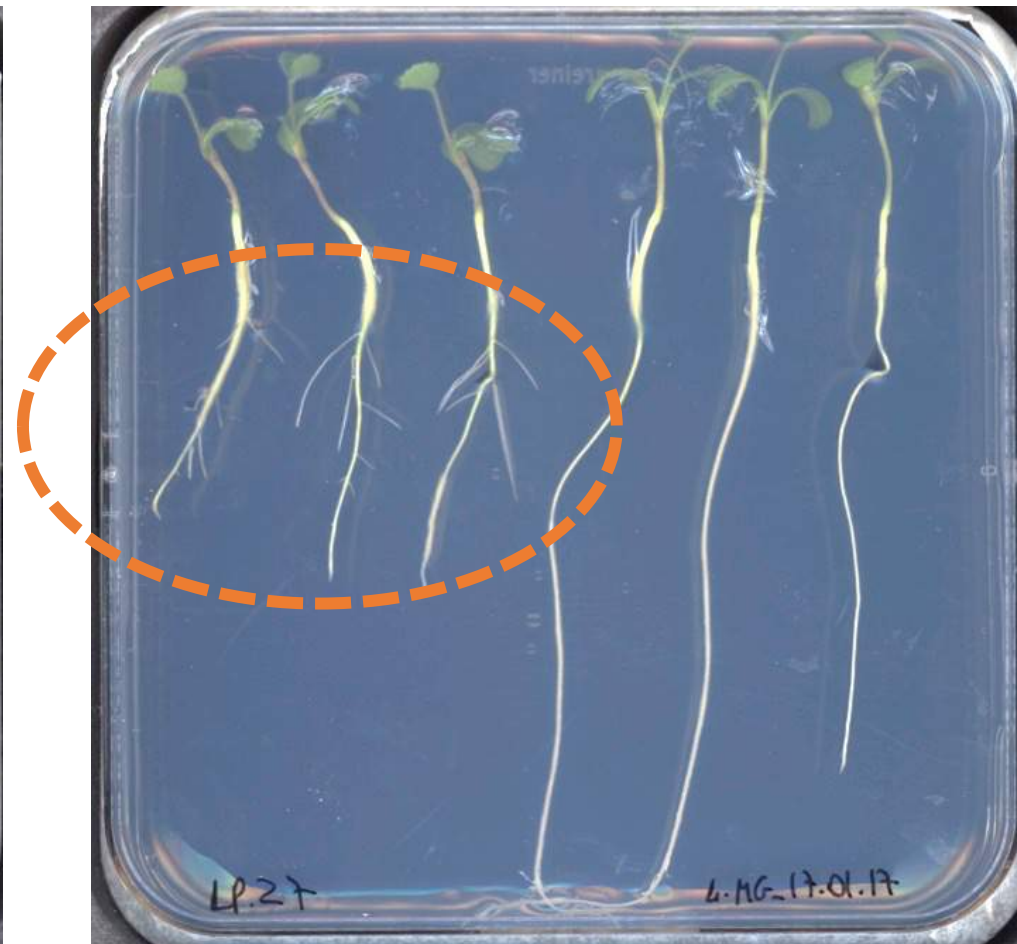


Low phosphate is increasing lateral root number exclusively on certain genotypes

phosphate

750 μM

20 μM



genotype

A

B

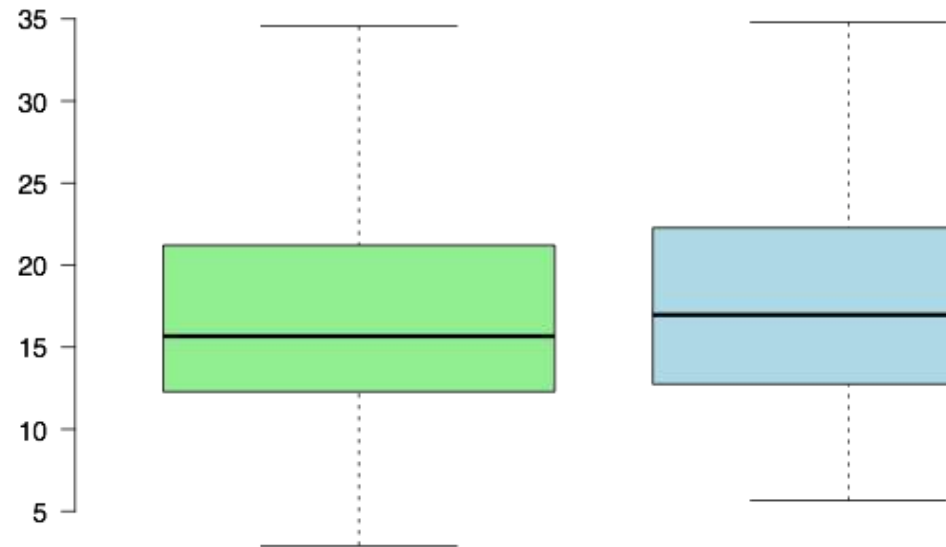
A

B

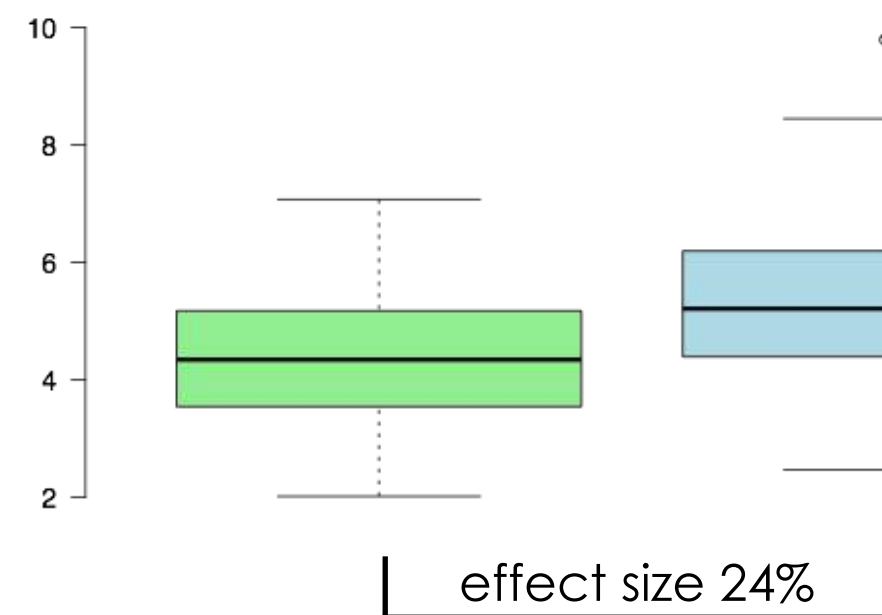
Day 7

Shoot anion concentration (ng/mg_{FW})

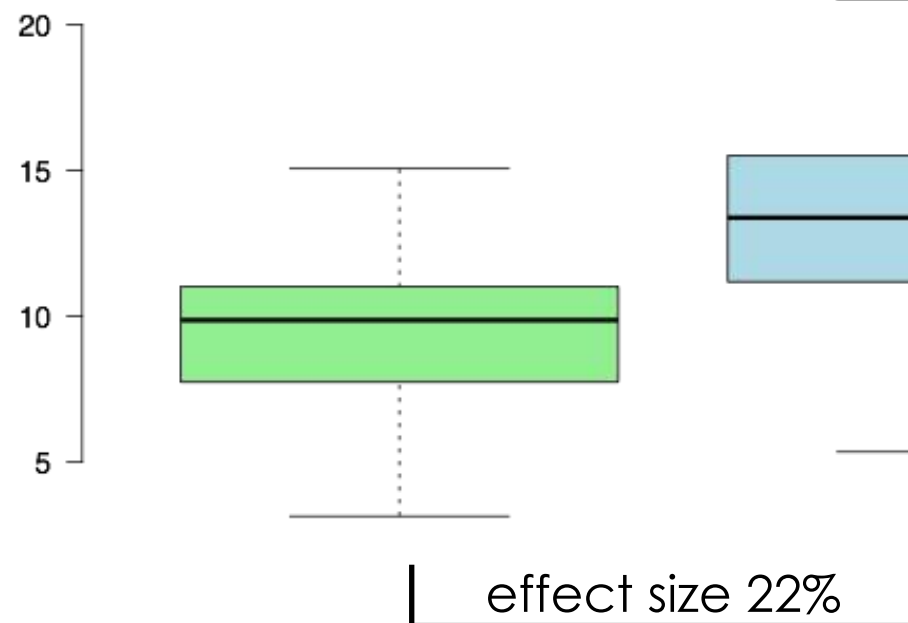
nitrate



sulfate



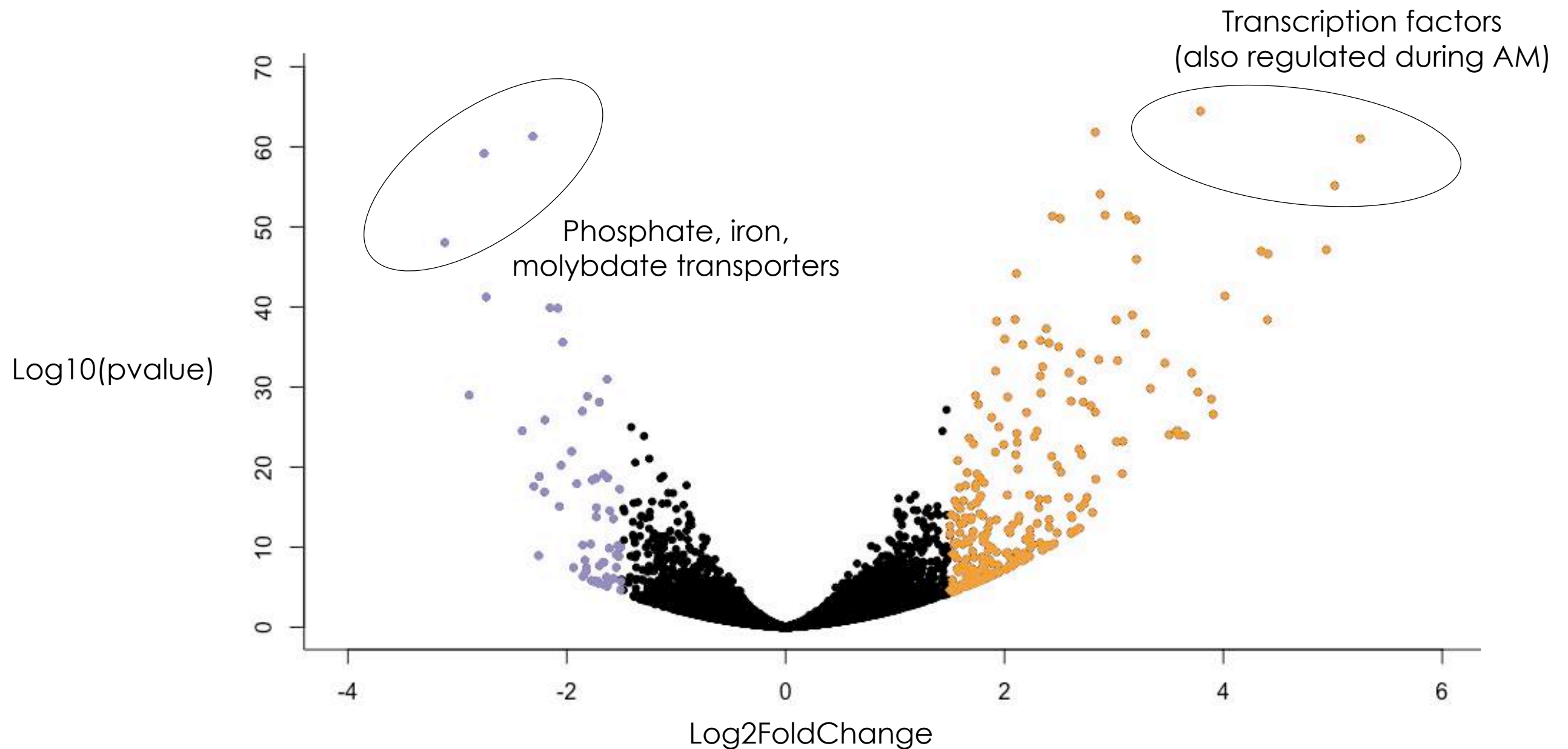
phosphate



20 μ M phosphate

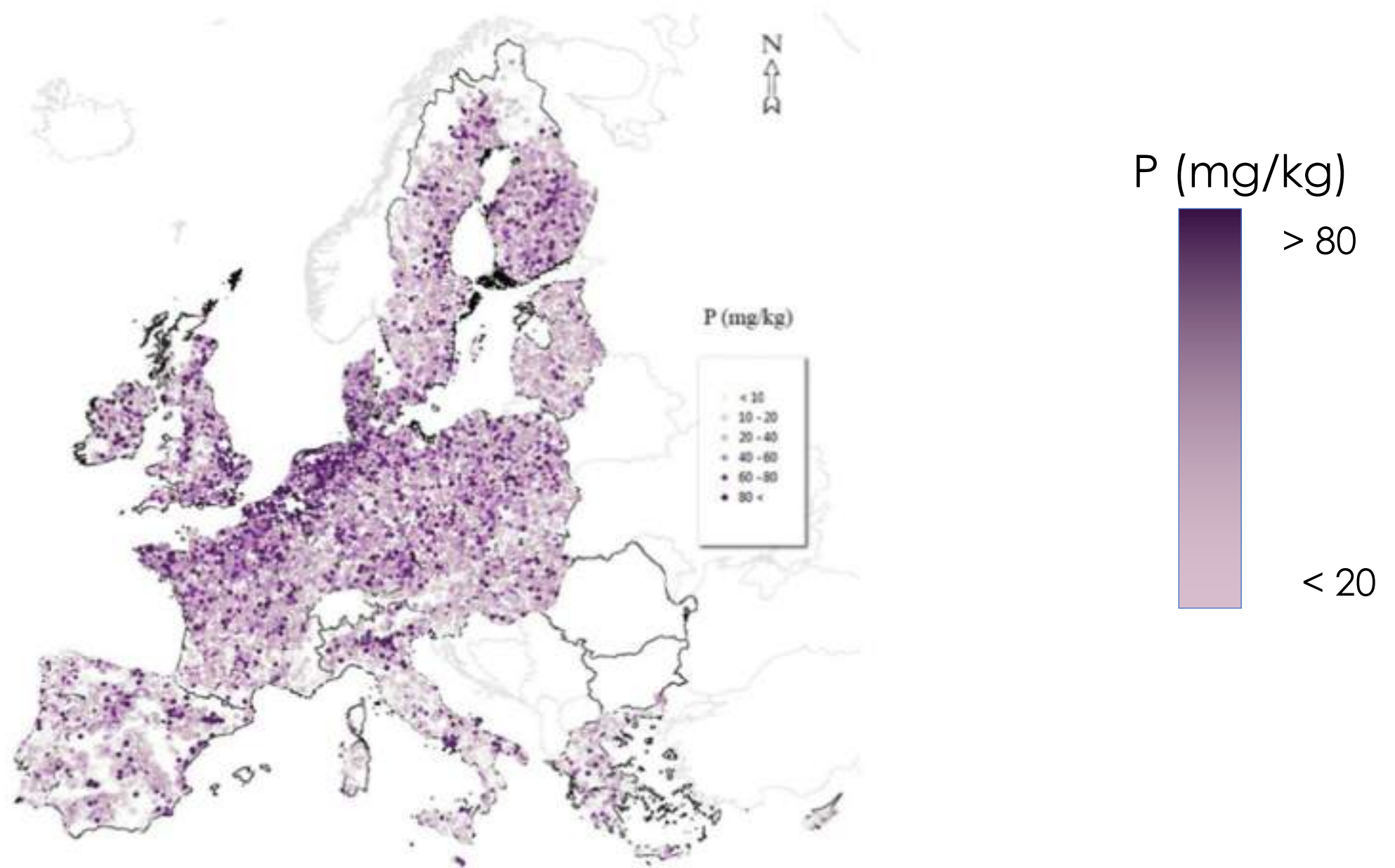
750 μ M phosphate

RNAseq reveals genes that are expressed in roots and influenced by phosphate levels



More than 60 **downregulated** and 285 **upregulated** genes
(Log2fold change >1.5 or <-1.5)
Perfect Atlas for following GWAS candidate genes

Top soluble phosphorous concentration



Different levels of phosphate are a selective pressure for specific alleles?

High Phosphate

$\text{Ca}(\text{NO}_3)_2$ 1.5 mM

KH_2PO_4 0.75 mM

KNO_3 1 mM

MgSO_4 0.75 mM

KCl -

Low Phosphate

$\text{Ca}(\text{NO}_3)_2$ 1.5 mM

KH_2PO_4 20 μM

KNO_3 1 mM

MgSO_4 0.75 mM

KCl 0.70 mM

+ 0.8% agarose
+ MES 0.8 g/L
pH 5.7

